**Uterine Malignant and Potentially Malignant Mesenchymal Tumours Histopathology Reporting Guide**

**Elements in black text are CORE Elements in grey text are NON-CORE o indicates single select values □ indicates multi-select values**

|  |  |
| --- | --- |
| Definition of Core elements | CORE elements are those which are essential for the clinical management, staging or prognosis of the cancer. These elements will either have evidentiary support at Level III-2 or above (based on prognostic factors in the National Health and Medical Research Council (NHMRC) levels of evidence1). In rare circumstances, where level III-2 evidence is not available an element may be made a CORE element where there is unanimous agreement by the Dataset Authoring Committee (DAC). An appropriate staging system e.g., Pathological TNM staging would normally be included as a CORE element.  Molecular and immunohistochemical testing is a growing feature of cancer reporting. However, in many parts of the world this type of testing is limited by the available resources. In order to encourage the global adoption of ancillary tests for patient benefit, International Collaboration on Cancer Reporting (ICCR) includes the most relevant ancillary testing in ICCR Datasets as CORE elements, especially when they are necessary for the diagnosis. Where the technical capability does not yet exist, laboratories may consider temporarily using these data elements as NON-CORE items.  The summation of all CORE elements is considered to be the minimum reporting standard for a specific cancer.  **Reference**  1 Merlin T, Weston A and Tooher R (2009). Extending an evidence hierarchy to include topics other than treatment: revising the Australian 'levels of evidence'. *BMC Med Res Methodol* 9:34. |
| Definition of Non-core elements | NON-CORE elements are those which are unanimously agreed should be included in the dataset but are not supported by level III-2 evidence. These elements may be clinically important and recommended as good practice but are not yet validated or regularly used in patient management.  Key information other than that which is essential for clinical management, staging or prognosis of the cancer such as macroscopic observations and interpretation, which are fundamental to the histological diagnosis and conclusion e.g., macroscopic tumour details, may be included as either CORE or NON-CORE elements by consensus of DAC. |
| Scope of this dataset | The dataset has been developed for the pathology reporting of resection specimens of the uterus for sarcomas and mesenchymal tumours with potentially malignant behaviour. The dataset is applicable to tumours of the uterine corpus and the uterine cervix.  Carcinomas, other non-mesenchymal malignancies and metastatic neoplasms are excluded from this dataset. Carcinosarcoma is also excluded as it is considered to represent a malignant epithelial tumour with divergent mesenchymal differentiation based on clinicopathologic, immunohistochemical and molecular analysis; as such, this entity is included in the ICCR Endometrial Cancer dataset. |

| **Core/**  **Non-core** | **Element name** | **Values** | **Commentary** | **Implementation notes** |
| --- | --- | --- | --- | --- |
| Non-core | CLINICAL INFORMATION | * Information not provided * History of previous cancer, *specify previous failed chemotherapy* * History of previous gynecologic biopsy/surgical excision, *specify* * Other, *specify* | Adequate clinical history is essential for accurate diagnoses and appropriate clinical care. It has been estimated that approximately 1% of diagnostic reports have been negatively impacted due to a lack of clinical information; in these instances, additional clinical information resulted in a change in diagnosis.1 A history of prior malignancy, radiation or hormonal therapy (which increases risk for sarcomas), and any prior excision are considered relevant.  **Reference**  1 Nakhleh RE, Gephardt G and Zarbo RJ (1999). Necessity of clinical information in surgical pathology. *Arch Pathol Lab Med* 123(7):615-619. |  |
| Core | OPERATIVE PROCEDURE | * Not specified * Hysterectomy * Simple total * Simple supracervical/subtotal * Radical * Type not specified * Myomectomy * Lymph nodes, *specify site(s)* * Other, *specify* | While a diagnosis of a uterine sarcoma or tumour of uncertain malignant potential may occur with limited sampling of disease (endometrial biopsy, curetting or core biopsy), a significant subset are clinically unsuspected and first diagnosed upon pathologic examination of a myomectomy or hysterectomy specimen. Hysterectomy, with or without bilateral salpingo-oophorectomy, and myomectomy can provide both diagnostic and complete surgical resection of disease, although myomectomy may be associated with residual tumour post-resection. Laparoscopic myomectomy/ hysterectomy followed by in vivo fragmentation (morcellation) affects specimen integrity, discussed below, and may be suboptimal for diagnosis because of distortion of the organ’s anatomy. In general, surgical management is related to tumour site and wish for fertility preservation and the decision to perform salpingo-oophorectomy depends on the disease type and the patient’s age as ovarian preservation in young patients with uterine sarcoma may not impact overall survival.1,2 Nevertheless, hysterectomy with or without bilateral salpingo-oophorectomy is the most common and complete type of resection for malignant mesenchymal tumours. Since some sarcomas more frequently metastasize to lymph nodes than others, planning lymph node sampling or dissection is partly based on the sarcoma type (if known preoperatively), presence of clinically evident nodal disease at time of surgery, and surgeon’s preference.  **References**  1 Nasioudis D, Mastroyannis SA, Latif NA, Ko EM, Haggerty AF, Kim SH, Morgan MA and Giuntoli RL, 2nd (2020). Effect of bilateral salpingo-oophorectomy on the overall survival of premenopausal patients with stage I low-grade endometrial stromal sarcoma; a National Cancer Database analysis. *Gynecol Oncol* 157(3):634-638.  2 Nasioudis D, Chapman-Davis E, Frey M and Holcomb K (2017). Safety of ovarian preservation in premenopausal women with stage I uterine sarcoma. *J Gynecol Oncol* 28(4):e46. |  |
| Core | SPECIMEN INTEGRITY | * Intact * Non-intact * Morcellated/fragmented * Other, *specify* | Documentation of specimen integrity is crucial for reporting of malignant and potentially malignant uterine mesenchymal tumours as integrity affects evaluation of margins and can impact staging and prognosis of uterine sarcomas.1-3 It is important to document morcellation, a surgical technique performed in vivo after laparoscopic myomectomy or hysterectomy to reduce the size of the specimen into fragments small enough to be removed from the patient through the laparoscopic incision sites. Recurrence of uterine sarcoma has been reported when tumours are removed laparoscopically with morcellation.4 After this phenomenon was first documented in several series,4-6 certain protective measures were encouraged by the gynaecology community regarding use of this particular surgical technique.7-9  **References**  1 Pautier P, Genestie C, Rey A, Morice P, Roche B, Lhommé C, Haie-Meder C and Duvillard P (2000). Analysis of clinicopathologic prognostic factors for 157 uterine sarcomas and evaluation of a grading score validated for soft tissue sarcoma. *Cancer* 88(6):1425-1431.  2 Gootee J, Sioda N, Aurit S, Curtin C and Silberstein P (2020). Important prognostic factors in leiomyosarcoma survival: a National Cancer Database (NCDB) analysis. *Clin Transl Oncol* 22(6):860-869.  3 Koivisto-Korander R, Butzow R, Koivisto AM and Leminen A (2008). Clinical outcome and prognostic factors in 100 cases of uterine sarcoma: experience in Helsinki University Central Hospital 1990-2001. *Gynecol Oncol* 111(1):74-81.  4 Seidman MA, Oduyebo T, Muto MG, Crum CP, Nucci MR and Quade BJ (2012). Peritoneal dissemination complicating morcellation of uterine mesenchymal neoplasms. *PLoS One* 7(11):e50058.  5 Oduyebo T, Rauh-Hain AJ, Meserve EE, Seidman MA, Hinchcliff E, George S, Quade B, Nucci MR, Del Carmen MG and Muto MG (2014). The value of re-exploration in patients with inadvertently morcellated uterine sarcoma. *Gynecol Oncol* 132(2):360-365.  6 George S, Barysauskas C, Serrano C, Oduyebo T, Rauh-Hain JA, Del Carmen MG, Demetri GD and Muto MG (2014). Retrospective cohort study evaluating the impact of intraperitoneal morcellation on outcomes of localized uterine leiomyosarcoma. *Cancer* 120(20):3154-3158.  7 Halaska MJ, Haidopoulos D, Guyon F, Morice P, Zapardiel I and Kesic V (2017). European Society of Gynecological Oncology Statement on Fibroid and Uterine Morcellation. *Int J Gynecol Cancer* 27(1):189-192.  8 Anonymous (2019). ACOG Committee Opinion No. 770: Uterine Morcellation for Presumed Leiomyomas. *Obstet Gynecol* 133(3):e238-e248.  9 Society of Gynecologic Oncology (2013). *SGO POSITION STATEMENT: MORCELLATION*. Available at: https://www.sgo.org/resources/morcellation/ (Accessed 17th February 2021). |  |
| Core | SPECIMEN(S) SUBMITTED | * None submitted * Ovaries * Left * Right * Not specified * Fallopian tubes * Left * Right * Not specified * Omentum * Peritoneal biopsies, *specify site(s)* * Peritoneal washings/peritoneal fluid * Lymph nodes, *specify site(s)* * Other, *specify* | The presence of accompanying organs or tissues, other than the primary tumour site specimen (myomectomy or hysterectomy) is important because it contributes to the pathological assessment of tumour extension, other than by imaging (see **EXTENT OF INVASION**) and staging. If peritoneal washings/peritoneal fluid are submitted, this should be documented along with the presence or absence of tumour cells (see **EXTENT OF INVASION**). |  |
| Core | TUMOUR SITE | * Indeterminate * Cervix * Lower uterine segment * Corpus * Other, *specify* | Uterine sarcomas can arise primarily in the cervix or corpus, and in most cases, the site can easily be assigned. If the origin of a sarcoma is equivocal and it is difficult to establish whether the tumour has arisen from the cervix or the corpus (including the isthmus), deference is typically given to a corpus origin.1 Some tumours cannot be assigned a site of origin, for example if they are removed piecemeal, such as with morcellation, or when they efface normal anatomy and/or present at high stage. In this instance, the ‘other’ category can be used with an explanatory note.  **Reference**  1 Fadare O (2006). Uncommon sarcomas of the uterine cervix: a review of selected entities. *Diagn Pathol* 1:30. |  |
| Core | MAXIMUM TUMOUR DIMENSION | * \_\_\_ mm * Cannot be assessed, *specify* | Maximum tumour measurement requires an intact tumour as dimensions cannot be assessed on piecemeal or morcellated specimens. As such, evaluation of this element usually requires a hysterectomy or myomectomy specimen. Tumour size, which is the gross measurement across the largest dimension, is given in millimetres, although International Federation of Gynecology and Obstetrics (FIGO) and TNM staging parameters require conversion to centimetres.  Measurement in three dimensions is not required, nevertheless, tumour size is an important quality measure. When a case is being reviewed, it allows the reviewing pathologist to assess whether the tumour has been ‘adequately’ sampled. This may be particularly important in tumours with variable or undifferentiated morphology.  Tumour size is also critical for staging and may have prognostic significance. Leiomyosarcomas and endometrial stromal sarcomas confined to the uterus and measuring less than 50 mm may have a more favourable prognosis, which is reflected in the staging (FIGO Stage IA versus IB),1 although some studies have shown no association between size and outcome for Stage I leiomyosarcoma.2 Size ≥50 mm is one of the parameters used to assess malignant potential in perivascular epithelioid cell tumours (PEComa) of gynaecological origin.3,4 For inflammatory myofibroblastic tumour (IMT), size >70 mm may be associated with an aggressive clinical course, although evidence is limited.5,6  **References**  1 Abeler VM, Røyne O, Thoresen S, Danielsen HE, Nesland JM and Kristensen GB (2009). Uterine sarcomas in Norway. A histopathological and prognostic survey of a total population from 1970 to 2000 including 419 patients. *Histopathology* 54(3):355-364.  2 Wang WL, Soslow R, Hensley M, Asad H, Zannoni GF, de Nictolis M, Branton P, Muzikansky A and Oliva E (2011). Histopathologic prognostic factors in stage I leiomyosarcoma of the uterus: a detailed analysis of 27 cases. *Am J Surg Pathol* 35(4):522-529.  3 Bennett JA, Braga AC, Pinto A, Van de Vijver K, Cornejo K, Pesci A, Zhang L, Morales-Oyarvide V, Kiyokawa T, Zannoni GF, Carlson J, Slavik T, Tornos C, Antonescu CR and Oliva E (2018). Uterine PEComas: A Morphologic, Immunohistochemical, and Molecular Analysis of 32 Tumors. *Am J Surg Pathol* 42(10):1370-1383.  4 Schoolmeester JK, Howitt BE, Hirsch MS, Dal Cin P, Quade BJ and Nucci MR (2014). Perivascular epithelioid cell neoplasm (PEComa) of the gynecologic tract: clinicopathologic and immunohistochemical characterization of 16 cases. *Am J Surg Pathol* 38(2):176-188.  5 Parra-Herran C, Quick CM, Howitt BE, Dal Cin P, Quade BJ and Nucci MR (2015). Inflammatory myofibroblastic tumor of the uterus: clinical and pathologic review of 10 cases including a subset with aggressive clinical course. *Am J Surg Pathol* 39(2):157-168.  6 Bennett JA, Nardi V, Rouzbahman M, Morales-Oyarvide V, Nielsen GP and Oliva E (2017). Inflammatory myofibroblastic tumor of the uterus: a clinicopathological, immunohistochemical, and molecular analysis of 13 cases highlighting their broad morphologic spectrum. *Mod Pathol* 30(10):1489-1503. |  |
| Non-core | BLOCK IDENTIFICATION KEY | List overleaf or separately with an indication of the nature  and origin of all tissue blocks | The origin/designation of all tissue blocks should be recorded. This information should ideally be documented in the final pathology report and is particularly important should the need for internal or external review arise. The reviewer needs to be clear about the origin of each block in order to provide an informed specialist opinion. If this information is not included in the final pathology report, it should be available on the laboratory computer system and relayed to the reviewing pathologist. It may be useful to have a digital image of the specimen and record of the origin of the tumour blocks in some cases.  Recording the origin/designation of tissue blocks also facilitates retrieval of blocks for further immunohistochemical or molecular analysis, research studies or clinical trials. |  |
| Core | HISTOLOGICAL TUMOUR TYPE | * Smooth muscle tumour of uncertain malignant potential (STUMP) * Leiomyosarcoma * Endometrial stromal sarcoma, low grade * Endometrial stromal sarcoma, high grade * Undifferentiated uterine sarcoma * Mullerian adenosarcoma without sarcomatous overgrowth * Mullerian adenosarcoma with sarcomatous overgrowth * Uterine tumour resembling ovarian sex cord tumour (UTROSCT) * Perivascular epithelioid cell tumour (PEComa) * Inflammatory myofibroblastic tumour * NTRK-rearranged sarcoma * SMARC-deficient uterine sarcoma * Rhabdomyosarcoma (RMS) (embryonal and pleomorphic) * Alveolar soft part sarcoma * Neuroendocrine neoplasm, *specify type* * Other, *specify* | Our knowledge of the different types of mesenchymal tumours that can occur in the uterus has expanded in the past decade, as underlying molecular abnormalities have helped define distinctive clinicopathologic entities. Proper classification of malignant and potentially malignant tumours is crucial as there are important differences in clinical management and outcome. In many instances, additional tumour sampling may be more useful than ancillary techniques; in particular, sampling of the border of tumours can be useful.  All mesenchymal tumours of the uterus should be typed according to the most recent edition of the World Health Organization (WHO) Classification of Tumours of [Female Genital Tumours](https://publications.iarc.fr/592), 5th edition, 2020 (Table 1).1The International Collaboration on Cancer Reporting dataset includes 5th edition Corrigenda, June 2021.2 The most commonly encountered sarcomas - leiomyosarcoma, endometrial stromal sarcoma and Müllerian adenosarcoma - will be discussed first.  **Table 1 (See end of the document for Table)**  Smooth muscle tumours  Classification of smooth muscle tumours primarily relies on histologic assessment of several parameters. This assessment, however, can be challenging as benign, malignant and tumours classified as of uncertain malignant potential can share overlapping morphologies. For example, a high degree of cellularity may be seen in both benign (cellular leiomyoma) and malignant (leiomyosarcoma) tumours. Nevertheless, using the most recent edition of the WHO Classification, most uterine smooth muscle neoplasms are readily diagnosed either as benign or malignant.4 The type of leiomyosarcoma (spindle, epithelioid, myxoid) should be included in the report.  Leiomyosarcoma with spindle cell differentiation is diagnosed when there are at least two of the following three histological parameters: diffuse, moderate to severe nuclear atypia, mitotic count ≥10 per 2 mm2 (≥10 mitoses per 10 high power fields (HPF) if field diameter is 0.55 mm) and tumour cell necrosis.24 The criteria for diagnosis of malignancy in epithelioid and myxoid smooth muscle tumours are stricter. Epithelioid leiomyosarcoma usually contains ≥4 mitoses per 2 mm2 (≥4 mitoses per 10 HPFs if field diameter is 0.55 mm) moderate to severe nuclear atypia and/or tumour cell necrosis.5-7 Myxoid leiomyosarcomas usually have an infiltrative border, and either moderate to severe nuclear atypia, tumour cell necrosis, or >1 mitosis per 2 mm2 (>1 mitoses per 10 HPFs if field diameter is 0.55 mm).8  Tumours which show morphological features that exceed the criteria for leiomyoma but fall below the threshold for leiomyosarcoma may be diagnosed as STUMP.9,10 The category of STUMP should be used sparingly and before making a diagnosis of STUMP, every effort should be made to establish a diagnosis of either a leiomyoma subtype, leiomyosarcoma, or one of the recently described mesenchymal tumours with deceptively bland cytology as included in this dataset (i.e., perivascular epithelioid cell tumour, IMT, and neurotrophic tyrosine receptor kinase (NTRK)-rearranged spindle cell sarcoma). Besides epithelioid and myxoid neoplasms, the most common histologic subtypes of leiomyoma which may give rise to diagnostic difficulties are leiomyoma with bizarre nuclei and cellular leiomyoma. In order to make a distinction from leiomyosarcoma, accurate assessment of the number of mitoses in leiomyoma with bizarre nuclei is important but this is not straightforward because karyorrhectic nuclei may mimic mitoses.11,12 Fumarate hydratase (FH)-deficient morphology can be seen in leiomyoma with bizarre nuclei as well as conventional and cellular leiomyomata. In FH-deficient leiomyomata, the nuclei are often arranged in chains, have eosinophilic cytoplasmic inclusions, prominent eosinophilic nuclei, and perinucleolar haloes. The presence of these features, often accompanied by staghorn blood vessels and alveolar-pattern edema, particularly in a smooth muscle tumour occurring in a young woman, should prompt consideration of association with fumarate hydratase deficiency. Loss of FH staining by immunohistochemistry supports the diagnosis. Of note, a subset of these tumours are characterised by intact expression of FH (corresponding to the presence of FH protein that is non-functional). With either loss or a non-functional FH protein, accumulation of 2-SC and a positive 2-SC stain confirms the diagnosis. Although the majority of these cases seems to be sporadic, hereditary leiomyoma and renal cell carcinoma syndrome needs to be ruled out in the appropriate clinical setting.13-16 As high cellularity may be observed in benign and malignant smooth muscle tumours, as well as endometrial stromal neoplasms, tumours considered cellular leiomyoma should not contain microscopic features which exceed the WHO criteria for leiomyoma.4  Smooth muscle tumour of uncertain malignant potential (STUMP) is an applicable diagnosis if a spindle cell smooth muscle tumour has focal/multifocal or diffuse nuclear atypia, 5-9 mitoses per 2 mm2 (5-9 mitoses per 10 HPFs if field diameter is 0.55 mm) and lacks tumour cell necrosis. Approximately 12-17% of such tumours have recurred. The STUMP diagnosis is also applicable to any bland smooth muscle tumour with tumour cell necrosis or necrosis of an uncertain type. Approximately 28% of such tumours have recurred. Tumours lacking cytological atypia and tumour cell necrosis, but with ≥15 mitoses per 2 mm2 (≥15 mitoses per 10 HPFs if field diameter is 0.55 mm) are also considered STUMPs. Although none of such cases has recurred, the experience with these tumours is limited.10,17 In addition to the Stanford criteria,17 other helpful parameters that may be included in the assessment of recurrent potential in smooth muscle neoplasms are atypical mitoses, vascular involvement, and infiltrative/irregular margins.18 Epithelioid and myxoid STUMPs are rare, and it is important to exclude their respective benign and malignant variants by integrating gross, microscopic and molecular findings.  Endometrial stromal sarcoma  The classification of endometrial stromal sarcoma has evolved over time due to a better understanding of its morphologic spectrum and underlying recurrent molecular abnormalities. While it was historically separated into low and high grade categories based on mitotic count, the category of high grade endometrial stromal sarcoma was removed from the 2003 WHO Classification as there was a lack of clinical relevance in separating tumours that morphologically resembled proliferative phase endometrial stroma into low and high grade categories based on mitotic count alone.19 It is worth noting that the category of high grade endometrial stromal sarcoma at that time represented a heterogeneous group of tumours including those that resembled endometrial stroma and those with more nuclear pleomorphism. Currently, two categories of endometrial stromal sarcoma are recognised by the most recent WHO Classification.1 While they maintain the same lexicon used in the past - low grade and high grade endometrial stromal sarcoma - they represent two distinct clinicopathologic entities with differing morphology, biologic behaviour and molecular findings.19-25  Low grade endometrial stromal sarcoma is composed of cells which morphologically resemble proliferative-phase endometrial stroma, i.e., cells have uniform round to ovoid nuclei and scant cytoplasm and they are associated with a delicate spiral arteriole-like network. This tumour has a characteristic growth pattern as it typically permeates the myometrium in a ‘finger-like’ or ‘tongue-like’ fashion; lymphovascular invasion (LVI) is frequent and sometimes prominent. Smooth muscle, sex cord-like, fibrous and myxoid variant morphology is not uncommon. Most, but not all tumours, harbour gene fusions most commonly *JAZF1-SUZ12.* Tumours with sex cord-like differentiation often harbour fusions involving *PHF1*. Patients with low grade endometrial stromal sarcoma typically have an indolent and protracted course.  Some low grade endometrial stromal tumours are classified as having ‘limited’ infiltration. These represent tumours that lack overt myometrial permeation but have more margin irregularity than allowed for designation as an endometrial stromal nodule.26,27 Although most behave in benign fashion, a subset of tumours classified as such have metastasized. Thus, these tumours should be regarded as potentially malignant and be classified as low grade endometrial stromal sarcomas with limited infiltration.28  High grade endometrial stromal sarcoma encompasses tumours that have distinctive clinical, histologic and molecular findings that differs from low grade endometrial stromal sarcoma.23,25,29-31 This tumour type occurs over a wide age range and shows a variable morphology but typically contains at least a focal characteristic round cell component (if associated with *YWHAE*-rearrangement), or myxoid spindle cell component (if associated with *BCOR*-rearrangement, most commonly *ZC3H7B-BCOR*, or internal tandem duplications). Patients with high grade endometrial stromal sarcoma more commonly present at higher stage in comparison to patients with low grade endometrial stromal sarcoma. Histologically, they can show expansile, permeative, or more commonly destructive infiltration of the myometrium; LVI can also be prominent. Tumours associated with *YWHAE*-rearrangement often, but not always, have a morphologically low grade component often akin to the fibromyxoid variant of low grade endometrial sarcoma. *BCOR*-associated tumours can closely mimic the appearance of myxoid leiomyosarcoma as tumour cells are often spindled with mild to moderate nuclear atypia and set in a prominent myxoid stroma. Limited clinical data suggest that high grade endometrial stromal sarcomas, regardless of the underlying genetic abnormality, are more likely to pursue an aggressive clinical course with earlier recurrences and metastasis, in comparison to low grade endometrial stromal sarcoma.23,25,29,31  Although rare, a scenario worth mentioning is the potential for low grade endometrial stromal sarcoma to ‘transform’ to a high grade tumour. In this scenario, the tumour may have the appearance of a high grade endometrial stromal sarcoma or undifferentiated uterine sarcoma but harbour translocations characteristic of conventional low grade endometrial stromal sarcoma.32  Müllerian adenosarcoma  Müllerian adenosarcoma is a biphasic neoplasm composed of a benign, non-neoplastic Müllerian epithelial component and a malignant sarcomatous component which is usually, but not always, morphologically low grade. These tumours are uncommon, representing less than 1% of all uterine malignancies and approximately 10% of uterine sarcomas. They present over a wide age range, most commonly in postmenopausal women, but a significant subset occurs in younger adults. Patients typically present with abnormal uterine bleeding. Other findings may include an enlarged uterus, pelvic mass, or polyp (either endocervical or endometrial in origin). Gross examination may show multiple large, soft polypoid masses filling the uterine cavity; tumours may invade the myometrium or cervical stroma, a finding more commonly associated with sarcomatous overgrowth. Characteristic histologic findings include a leaf-like growth pattern (also often described as phyllodes-like as the appearance is akin to a phyllodes tumour of the breast), with intraglandular stromal polypoid projections, and cuffing of the glands by hypercellular stroma. However, not all adenosarcomas show phylloidiform growth with some being composed of variably sized rounded glands surrounded by hypercellular stroma (‘rigid cysts’); a combination of these appearances is not uncommon. The stromal cells may show variable amounts of nuclear atypia in the form of nuclear enlargement with irregular nuclear contour or nuclear hyperchromasia. Mitoses are typically identified (usually >1 per 2 mm2 (>1 mitosis per 10 HPFs if field diameter is 0.55 mm)) but may be sparse or, in rare cases, absent. The stromal component is most commonly homologous, i.e., it has the appearance of endometrial or cervical stroma, but may also show heterologous differentiation, most commonly rhabdomyosarcoma. Sex cord-like differentiation may also occur. Sarcomatous overgrowth is defined as the presence of greater than 25% of the tumour composed solely of a neoplastic stromal component without epithelium; sex-cord-like differentiation is not considered in the assessment of stromal overgrowth.33,34 Sarcomatous overgrowth often shows aberrant p53 immunoreactivity and loss of hormone receptor positivity.35,36 It is important to note that the non-neoplastic epithelial component typically has a banal appearance, sometimes with various types of epithelia (tubal, endometrioid, mucinous, squamous); occasionally the epithelial component may show some cytologic atypia in the form of nuclear enlargement and hyperchromasia. In this latter scenario, additional sampling may be prudent to exclude carcinosarcoma. Features associated with an unfavourable outcome include sarcomatous overgrowth, deep myometrial invasion, and extrauterine extension; morphologically high grade nuclear atypia (marked nuclear enlargement and hyperchromasia) that shows mutation-type staining pattern for p53 may also be an adverse prognostic feature.36  Undifferentiated uterine sarcomas (Unclassifiable sarcomas)  Sarcomas which cannot be classified are considered undifferentiated uterine sarcoma. This is a diagnosis of exclusion after other malignancies, such as undifferentiated carcinoma, carcinosarcoma, leiomyosarcoma, and high grade endometrial stromal sarcoma have been excluded. As many of the tumours in the differential diagnosis may have areas of morphologic overlap, the diagnosis of undifferentiated uterine sarcoma is best rendered on a complete excision specimen as a more limited specimen may lack the diagnostic features of other uterine malignancies. It is worth noting that undifferentiated uterine sarcoma can be separated into two different types based on their morphologic appearance: uniform and pleomorphic.37 With advanced molecular techniques and newly reported molecular abnormalities, the former are increasingly categorised as high grade endometrial stromal sarcomas whereas the pleomorphic type likely mostly represent sarcomas that are so poorly differentiated that cannot be classified.38 Some of these tumours may represent carcinosarcomas in which only the sarcomatous component is seen, and thus before making a diagnosis of an undifferentiated uterine sarcoma, additional sampling should be considered which may reveal diagnostic areas. Prior to rendering the diagnosis of undifferentiated uterine sarcoma in an excision specimen, extensive tumour sampling, immunohistochemical staining, and if possible, molecular testing may be needed to exclude other neoplasms.  Potentially malignant mesenchymal tumours of the uterus are those in which prognostication requires assessment of various clinical and pathologic parameters to determine biologic potential. Tumours within this category include uterine tumour resembling ovarian sex cord tumour, perivascular epithelioid cell tumour, and IMT.  Uterine tumour resembling ovarian sex cord tumour (UTROSCT)  Uterine tumour resembling ovarian sex cord tumour (UTROSCT) is an uncommon uterine tumour whose histologic features recapitulates the appearance of an ovarian sex cord tumour. Historically, the term UTROSCT included tumours entirely composed of sex cord elements as well as endometrial stromal tumours with extensive sex cord differentiation; the latter are no longer considered in this category based on morphologic as well as molecular differences. UTROSCT exhibit a wide range of morphologic appearances with diffuse, corded, trabecular, tubular, retiform and/or nested growth. Tumour cells have variable amounts of cytoplasm ranging from inconspicuous to abundant, which may be pale, foamy or eosinophilic; rhabdoid morphology may be seen and can be extensive. Nuclei are usually uniform with minimal cytologic atypia and the mitotic count is low. Nuclear atypia in the form of nuclear enlargement and hyperchromasia, as well as brisk mitotic activity, may be seen. UTROSCT is characterised by recurrent gene fusions involving *NCOA1-3*, *GREB1* and *ESR1*.39-42 These tumours are considered to be of uncertain malignant potential. Although data is limited, features that may be associated with aggressive behaviour include a mitotic count >2 per 2 mm2 (>2 mitoses per 10 HPFs if field diameter is 0.55 mm), necrosis, extensive (>50%) rhabdoid morphology and potentially tumours with *GREB1* rearrangement.40-43  Perivascular epithelioid cell tumour (PEComa)  Perivascular epithelioid cell tumours (PEComas) are unusual mesenchymal neoplasms that are composed of a distinctive population of cells, termed perivascular epithelioid cells, which co-express smooth muscle and melanocytic markers. These tumours have wide anatomic distribution and the uterus is the most common site when they occur in the female genital tract.44-48 Most tumours occur sporadically with only a small subset being associated with tuberous sclerosis. Histologically, tumours most commonly are composed of epithelioid and spindle cells but can sometimes be solely or predominantly epithelioid or spindled. Tumours often, but not always, show a characteristic perivascular pattern of growth in which the tumour cells are radially arranged around the vasculature. In some tumours, the neoplastic cells can be seen within the muscular wall of the vessel. Another distinctive aspect of these tumours is their cytologic appearance; the cells are remarkable for abundant granular eosinophilic or clear cytoplasm although predominantly spindled tumours may show less abundant cytoplasm. When epithelioid, the tumour cells grow in sheets, nests and/or trabeculae that are surrounded by a delicate capillary vasculature. Spindled tumours often exhibit fascicular growth and can mimic smooth muscle neoplasia, the distinctive morphologic difference being the granular appearance of the cytoplasm. Of note, *TFE3*-associated PEComas often are composed of epithelioid cells that have predominantly clear cytoplasm; extensive melanin deposition may also occur. Assessment of potential malignant behaviour for PEComa of the female genital tract is based on the following parameters: tumour size ≥50 mm, high nuclear grade, mitotic count of >1 mitosis per 12 mm2, presence of necrosis and presence of vascular invasion. If a tumour has three or more of these features, it is best classified as malignant. Only tumours that lack all features could potentially be considered benign. Any tumour with one or two features should be considered of uncertain malignant potential.  Inflammatory myofibroblastic tumour (IMT)  Inflammatory myofibroblastic tumour (IMT) is a myofibroblastic/fibroblastic neoplasm characterised by a variably myxoid stroma with an accompanying variably intense inflammatory infiltrate, primarily composed of lymphocytes and plasma cells. This tumour shows a wide anatomic distribution with the uterine corpus being the most common location in the female genital tract; less commonly it involves the cervix.49-52 Occasionally, IMT may be identified at the time of delivery and in some cases may be adherent to the maternal surface of the placenta or be associated with the placental membranes.53,54 Microscopically, the tumour borders can be well demarcated or irregular, either showing permeative (stromal sarcoma-like) or infiltrative margins. A number of different morphologies may be seen and are often intermixed: myxoid, leiomyoma-like, or hyalinised. The myxoid pattern is characteristically hypocellular with individual cells dispersed in an abundant myxoid matrix, a feature that imparts a fasciitis-like appearance on low power magnification. The leiomyoma-like areas are composed of spindled cells in intersecting fascicles or showing storiform growth; the former closely mimics smooth muscle neoplasia. The hyalinised pattern is remarkable for an abundant hyalinised and collagenous stroma containing scattered spindled cells. The tumour cells in the fasciitis-like areas are spindled with eosinophilic to amphophilic cytoplasmic processes and ovoid to tapered nuclei with open dispersed chromatin, features that closely resemble the appearance of the spindled cells of nodular fasciitis. The spindled cells in the leiomyoma-like areas have features indistinguishable from smooth muscle neoplasia with eosinophilic cytoplasm and more oblong nuclei with blunt ends. The epithelioid variant of IMT, which typically occurs in the abdominal cavity and is characterised by a predominant component of epithelioid cells with eosinophilic cytoplasm and vesicular nuclei, has yet to be described in the uterus, but has been reported in the ovary.55,56 An inflammatory infiltrate, often present at the periphery of the tumour but also dispersed throughout, is typically composed of lymphocytes and plasma cells although other inflammatory cells can be seen; the amount and distribution of inflammatory cells vary but inflammation is typically a reproducible finding and characteristic of IMT. Assessment of potential malignant behaviour for IMT of the female genital tract is not well established. Some tumours present at high stage and should be considered malignant. Pathologic features which have been associated with aggressive behaviour include large tumour size (>150 mm), marked nuclear atypia, LVI and tumours with high mitotic counts (>10 per 2 mm2 which corresponds to >10 mitoses per 10 HPFs if field diameter is 0.55 mm). These features are not invariably associated with adverse outcome and conversely tumours as small as 62 mm or with mitotic counts of only 1 per 2 mm2 (1 mitosis per 10 HPFs if field diameter is 0.55 mm) have recurred. Recurrence of IMT at all anatomic sites is estimated at 25% and is related to resectability.57 As complete resection is typically achieved with hysterectomy, this may partially explain the overall good outcome for most patients with uterine neoplasms.  Rhabdomyosarcoma  Less common uterine sarcomas include rhabdomyosarcoma and alveolar soft part sarcoma. Different types of rhabdomyosarcoma have been described in the female genital tract; in the uterus, embryonal rhabdomyosarcoma (ERMS) and pleomorphic rhabdomyosarcoma are the most likely to be encountered.58-61 Histologically, ERMS characteristically shows alternating cellularity with hypocellular myxoid zones and hypercellular foci of spindled rhabdomyoblasts, often condensing underneath the overlying epithelium (cambium layer). Heterologous cartilaginous differentiation is commonly seen. Pleomorphic rhabdomyosarcoma is composed of sheets of highly atypical spindled, polygonal or rhabdoid cells with large irregular, frequently multinucleated cells and eosinophilic cytoplasm. Mitoses are frequent and often atypical. Both subtypes of rhabdomyosarcoma are positive for desmin, myoD1 and myogenin. ERMS is associated with high frequency of *DICER1* mutation which may be somatic or germline. Adult patients with ERMS have a less favourable prognosis than children.58 Patients with pleomorphic rhabdomyosarcoma have a poor prognosis.61 Some of these tumours may represent rhabdomyosarcomatous overgrowth in an adenosarcoma or carcinosarcoma; thus, extensive sampling should be undertaken to exclude an epithelial component before diagnosing a pleomorphic rhabdomyosarcoma.  Alveolar soft part sarcoma (ASPS)  Alveolar soft part sarcoma (ASPS) can occur at any location in the female genital tract; uterine tumours can occur in the corpus or cervix.62-65 Histologically, they are composed of nests of large, polygonal epithelioid cells with abundant eosinophilic granular cytoplasm containing an eccentric or centrally located nucleus with vesicular nuclei and prominent nucleolus. Characteristically, the nests are enveloped by a delicate sinusoidal vascular network and the cells often show dyscohesion resulting in an alveolar-like appearance. Cytoplasmic clearing and rhabdoid cells may be seen. Intracytoplasmic granules and/or rhomboid crystals may be seen, which can be highlighted by periodic acid-Schiff stain and are diastase resistant. Mitoses are typically sparse. ASPS is characterised by *TFE3* rearrangements as a result of chromosomal translocation t(x;17)(p11;q25). As a consequence, tumour cells typically show strong and diffuse nuclear staining for TFE3. The prognosis of ASPS at all anatomic sites appears to be related to resectability, which may explain the relative better prognosis for ASPS of the gynaecologic tract in comparison to those that arise elsewhere.  It is worth noting that many different types of sarcoma more commonly encountered in other anatomic locations can also rarely arise in the uterus, such as liposarcoma66 and angiosarcoma.67-69 Prior to making the diagnosis of a pure unusual type of sarcoma of the uterus, additional sampling of the lesion should be performed to exclude the possibility that it represents the component of a more commonly encountered uterine neoplasm, such as sarcomatous overgrowth of an adenosarcoma or the mesenchymal component of a carcinosarcoma.  Emerging entities  Emerging uterine mesenchymal entities include *NTRK*-rearranged sarcoma, *PDGFR*-rearranged sarcoma and *SMARCA4*-deficient uterine sarcoma. *NTRK*-rearranged sarcoma has been recently described to occur in the uterine cervix and lower uterine segment.70-73 Histologically, *NTRK*-rearranged sarcomas typically have an infiltrative border and are composed of a proliferation of spindled cells exhibiting either a patternless architecture or showing (often haphazard) fascicular or herringbone growth. Entrapped endocervical glands may be encircled by the neoplastic cells, sometimes with polypoid projections simulating adenosarcoma; however, there is typically no periglandular stromal condensation. The spindle cells have eosinophilic cytoplasm and generally show mild to moderate nuclear enlargement with nuclei that are ovoid with dispersed chromatin and small nucleoli; epithelioid change and foci of marked atypia may be seen. The vascular component can be composed of delicate capillaries or vessels with variably thickened walls often with prominent hyalinisation. The mitotic count is variable ranging from 0 to 50 per 2 mm2 (0-50 mitoses per 10 HPFs if field diameter is 0.55 mm) atypical mitotic figures and necrosis may be seen. Other findings that may be encountered include focal myxoid matrix, focal hemangiopericytoma-like vasculature and a prominent lymphocytic infiltrate. These tumours show positivity for pan-TRK, but this marker is not specific for the gene fusion. Patients with *NTRK*-rearranged sarcoma typically present with Stage I disease; however, approximately one third have developed recurrence or metastatic disease.70-73 Targeted therapy against tropomyosine kinase receptors has shown clinical benefit in patients with *NTRK*-associated sarcomas.74  *RET* fusion positive neoplasms may also exhibit fibroblastic or neural-like differentiation and have phenotypic overlap with *NTRK*-related neoplasms.75 Recently a cervical sarcoma with a novel *RET-SPECC1L* fusion has been described.76 Rare spindle cell sarcomas with recurrent *MEIS1-NCOA2* fusions have also been recently described.77  *COL1A1-PDGFB* rearranged uterine sarcomas are rare and data is limited.72,78 These tumours are composed of a cellular proliferation of spindle cells that typically exhibit a storiform or herringbone growth pattern although one tumour has been described as showing a fascicular ‘leiomyoma-like’ growth pattern. Overall, it is interesting to speculate that these tumours could be the uterine counterpart of dermatofibrosarcoma protuberans, as they show morphological overlap (including fibrosarcomatous areas), a similar immunophenotype (focal loss of CD34 staining in ‘fibrosarcomatous areas’), as well as sharing the same gene fusion.  *SMARCA4*-deficient uterine sarcoma (SDUS) is a recently described entity that shares morphologic overlap with undifferentiated endometrial carcinoma but has distinctive clinicopathologic and molecular differences.79-81 Tumours characteristically show a diffuse growth of large epithelioid cells with round vesicular nuclei and exhibit prominent rhabdoid morphology; other features which may be focally present include phyllodiform architecture, vague cording or nesting associated with stromal hyalinisation, small cell or spindled morphology, and focal myxoid stromal change. Brisk mitoses (usually >20 per 10 HPFs/0.24 mm2), necrosis and LVI are common. Patients with SDUS have a poor prognosis.79-81  **References**  1 WHO Classification of Tumours Editorial Board (2020). *Female Genital Tumours, WHO Classification of Tumours, 5th Edition, Volume 4*. IARC Press, Lyon.  2 WHO Classification of Tumours Editorial Board (2021). *Female Genital Tumours, WHO Classification of Tumours, 5th Edition, Volume 4 - Corrigenda June 2021*. Available from: https://publications.iarc.fr/Book-And-Report-Series/Who-Classification-Of-Tumours/Female-Genital-Tumours-2020 (Accessed 16th June 2021).  3 Fritz A, Percy C, Jack A,  Shanmugaratnam K, Sobin L, Parkin DM  and Whelan S (eds) (2020). *International Classification of Diseases for Oncology, Third edition, Second revision ICD-O-3.2*. Available from: http://www.iacr.com.fr/index.php?option=com\_content&view=category&layout=blog&id=100&Itemid=577 (Accessed 16th June 2021).  4 Ip PPC et al (eds) (2020). Uterine leiomyoma. In: *Female Genital Tumours, WHO Classification of Tumours, 5th Edition, Volume 4*, WHO Classification of Tumours Editorial Board (ed), IARC Press, Lyon.  5 Prayson RA, Goldblum JR and Hart WR (1997). Epithelioid smooth-muscle tumors of the uterus: a clinicopathologic study of 18 patients. *Am J Surg Pathol* 21(4):383-391.  6 Kurman RJ and Norris HJ (1976). Mesenchymal tumors of the uterus. VI. Epithelioid smooth muscle tumors including leiomyoblastoma and clear-cell leiomyoma: a clinical and pathologic analysis of 26 cases. *Cancer* 37(4):1853-1865.  7 Oliva E (2016). Practical issues in uterine pathology from banal to bewildering: the remarkable spectrum of smooth muscle neoplasia. *Mod Pathol* 29 Suppl 1:S104-120.  8 Parra-Herran C, Schoolmeester JK, Yuan L, Dal Cin P, Fletcher CD, Quade BJ and Nucci MR (2016). Myxoid Leiomyosarcoma of the Uterus: A Clinicopathologic Analysis of 30 Cases and Review of the Literature With Reappraisal of Its Distinction From Other Uterine Myxoid Mesenchymal Neoplasms. *Am J Surg Pathol* 40(3):285-301.  9 Ip PP, Cheung AN and Clement PB (2009). Uterine smooth muscle tumors of uncertain malignant potential (STUMP): a clinicopathologic analysis of 16 cases. *Am J Surg Pathol* 33(7):992-1005.  10 Ip PPC, Croce S and Gupta M (eds) (2020). Smooth muscle tumour of uncertain malignant potential of the uterine corpus. In: *Female Genital Tumours, WHO Classification of Tumours, 5th Edition, Volume 4*, WHO Classification of Tumours Editorial Board (ed), IARC Press, Lyon.  11 Bennett JA, Weigelt B, Chiang S, Selenica P, Chen YB, Bialik A, Bi R, Schultheis AM, Lim RS, Ng CKY, Morales-Oyarvide V, Young RH, Reuter VE, Soslow RA and Oliva E (2017). Leiomyoma with bizarre nuclei: a morphological, immunohistochemical and molecular analysis of 31 cases. *Mod Pathol* 30(10):1476-1488.  12 Chow KL, Tse KY, Cheung CL, Wong KW, Cheung AN, Wong RW, Chan AN, Yuen NW, Ngan HY and Ip PP (2017). The mitosis-specific marker phosphohistone-H3 (PHH3) is an independent prognosticator in uterine smooth muscle tumours: an outcome-based study. *Histopathology* 70(5):746-755.  13 Joseph NM, Solomon DA, Frizzell N, Rabban JT, Zaloudek C and Garg K (2015). Morphology and Immunohistochemistry for 2SC and FH Aid in Detection of Fumarate Hydratase Gene Aberrations in Uterine Leiomyomas From Young Patients. *Am J Surg Pathol* 39(11):1529-1539.  14 Sanz-Ortega J, Vocke C, Stratton P, Linehan WM and Merino MJ (2013). Morphologic and molecular characteristics of uterine leiomyomas in hereditary leiomyomatosis and renal cancer (HLRCC) syndrome. *Am J Surg Pathol* 37(1):74-80.  15 Miettinen M, Felisiak-Golabek A, Wasag B, Chmara M, Wang Z, Butzow R and Lasota J (2016). Fumarase-deficient Uterine Leiomyomas: An Immunohistochemical, Molecular Genetic, and Clinicopathologic Study of 86 Cases. *Am J Surg Pathol* 40(12):1661-1669.  16 Reyes C, Karamurzin Y, Frizzell N, Garg K, Nonaka D, Chen YB and Soslow RA (2014). Uterine smooth muscle tumors with features suggesting fumarate hydratase aberration: detailed morphologic analysis and correlation with S-(2-succino)-cysteine immunohistochemistry. *Mod Pathol* 27(7):1020-1027.  17 Bell SW, Kempson RL and Hendrickson MR (1994). Problematic uterine smooth muscle neoplasms. A clinicopathologic study of 213 cases. *Am J Surg Pathol* 18(6):535-558.  18 Gupta M, Laury AL, Nucci MR and Quade BJ (2018). Predictors of adverse outcome in uterine smooth muscle tumours of uncertain malignant potential (STUMP): a clinicopathological analysis of 22 cases with a proposal for the inclusion of additional histological parameters. *Histopathology* 73(2):284-298.  19 Chang KL, Crabtree GS, Lim-Tan SK, Kempson RL and Hendrickson MR (1990). Primary uterine endometrial stromal neoplasms. A clinicopathologic study of 117 cases. *Am J Surg Pathol* 14(5):415-438.  20 Koontz JI, Soreng AL, Nucci M, Kuo FC, Pauwels P, van Den Berghe H, Dal Cin P, Fletcher JA and Sklar J (2001). Frequent fusion of the JAZF1 and JJAZ1 genes in endometrial stromal tumors. *Proc Natl Acad Sci U S A* 98(11):6348-6353.  21 Nucci MR, Harburger D, Koontz J, Dal Cin P and Sklar J (2007). Molecular analysis of the JAZF1-JJAZ1 gene fusion by RT-PCR and fluorescence in situ hybridization in endometrial stromal neoplasms. *Am J Surg Pathol* 31(1):65-70.  22 Huang HY, Ladanyi M and Soslow RA (2004). Molecular detection of JAZF1-JJAZ1 gene fusion in endometrial stromal neoplasms with classic and variant histology: evidence for genetic heterogeneity. *Am J Surg Pathol* 28(2):224-232.  23 Lee CH, Mariño-Enriquez A, Ou W, Zhu M, Ali RH, Chiang S, Amant F, Gilks CB, van de Rijn M, Oliva E, Debiec-Rychter M, Dal Cin P, Fletcher JA and Nucci MR (2012). The clinicopathologic features of YWHAE-FAM22 endometrial stromal sarcomas: a histologically high-grade and clinically aggressive tumor. *Am J Surg Pathol* 36(5):641-653.  24 Hoang LN, Aneja A, Conlon N, Delair DF, Middha S, Benayed R, Hensley ML, Park KJ, Hollmann TJ, Hameed MR, Antonescu CR, Soslow RA and Chiang S (2017). Novel High-grade Endometrial Stromal Sarcoma: A Morphologic Mimicker of Myxoid Leiomyosarcoma. *Am J Surg Pathol* 41(1):12-24.  25 Lewis N, Soslow RA, Delair DF, Park KJ, Murali R, Hollmann TJ, Davidson B, Micci F, Panagopoulos I, Hoang LN, Arias-Stella JA, 3rd, Oliva E, Young RH, Hensley ML, Leitao MM, Jr., Hameed M, Benayed R, Ladanyi M, Frosina D, Jungbluth AA, Antonescu CR and Chiang S (2018). ZC3H7B-BCOR high-grade endometrial stromal sarcomas: a report of 17 cases of a newly defined entity. *Mod Pathol* 31(4):674-684.  26 Tavassoli FA and Norris HJ (1981). Mesenchymal tumours of the uterus. VII. A clinicopathological study of 60 endometrial stromal nodules. *Histopathology* 5(1):1-10.  27 Dionigi A, Oliva E, Clement PB and Young RH (2002). Endometrial stromal nodules and endometrial stromal tumors with limited infiltration: a clinicopathologic study of 50 cases. *Am J Surg Pathol* 26(5):567-581.  28 Moore M and McCluggage WG (2020). Uterine Endometrial Stromal Tumors With Limited Infiltration: First Report of a Case Series Indicating Potential for Malignant Behavior. *Int J Gynecol Pathol* 39(3):221-226.  29 Mariño-Enriquez A, Lauria A, Przybyl J, Ng TL, Kowalewska M, Debiec-Rychter M, Ganesan R, Sumathi V, George S, McCluggage WG, Nucci MR, Lee CH and Fletcher JA (2018). BCOR Internal Tandem Duplication in High-grade Uterine Sarcomas. *Am J Surg Pathol* 42(3):335-341.  30 Hoang L, Chiang S and Lee CH (2018). Endometrial stromal sarcomas and related neoplasms: new developments and diagnostic considerations. *Pathology* 50(2):162-177.  31 Micci F, Heim S and Panagopoulos I (2021). Molecular pathogenesis and prognostication of "low-grade'' and "high-grade" endometrial stromal sarcoma. *Genes Chromosomes Cancer* 60(3):160-167.  32 Zou Y, Turashvili G, Soslow RA, Park KJ, Croce S, McCluggage WG, Stewart CJR, Oda Y, Oliva E, Young RH, Da Cruz Paula A, Dessources K, Ashley CW, Hensley ML, Yip S, Weigelt B, Benayed R, Antonescu CR, Lee CH and Chiang S (2020). High-grade transformation of low-grade endometrial stromal sarcomas lacking YWHAE and BCOR genetic abnormalities. *Mod Pathol* 33(9):1861-1870.  33 Clement PB and Scully RE (1989). Müllerian adenosarcomas of the uterus with sex cord-like elements. A clinicopathologic analysis of eight cases. *Am J Clin Pathol* 91(6):664-672.  34 Stolnicu S, Molnar C, Barsan I, Boros M, Nogales FF and Soslow RA (2016). The Impact on Survival of an Extensive Sex Cord-like Component in Mullerian Adenosarcomas: A Study Comprising 6 Cases. *Int J Gynecol Pathol* 35(2):147-152.  35 Blom R and Guerrieri C (1999). Adenosarcoma of the uterus: a clinicopathologic, DNA flow cytometric, p53 and mdm-2 analysis of 11 cases. *Int J Gynecol Cancer* 9(1):37-43.  36 Hodgson A, Amemiya Y, Seth A, Djordjevic B and Parra-Herran C (2017). High-grade Müllerian Adenosarcoma: Genomic and Clinicopathologic Characterization of a Distinct Neoplasm With Prevalent TP53 Pathway Alterations and Aggressive Behavior. *Am J Surg Pathol* 41(11):1513-1522.  37 Kurihara S, Oda Y, Ohishi Y, Iwasa A, Takahira T, Kaneki E, Kobayashi H, Wake N and Tsuneyoshi M (2008). Endometrial stromal sarcomas and related high-grade sarcomas: immunohistochemical and molecular genetic study of 31 cases. *Am J Surg Pathol* 32(8):1228-1238.  38 Cotzia P, Benayed R, Mullaney K, Oliva E, Felix A, Ferreira J, Soslow RA, Antonescu CR, Ladanyi M and Chiang S (2019). Undifferentiated Uterine Sarcomas Represent Under-Recognized High-grade Endometrial Stromal Sarcomas. *Am J Surg Pathol* 43(5):662-669.  39 Dickson BC, Childs TJ, Colgan TJ, Sung YS, Swanson D, Zhang L and Antonescu CR (2019). Uterine Tumor Resembling Ovarian Sex Cord Tumor: A Distinct Entity Characterized by Recurrent NCOA2/3 Gene Fusions. *Am J Surg Pathol* 43(2):178-186.  40 Bennett JA, Lastra RR, Barroeta JE, Parilla M, Galbo F, Wanjari P, Young RH, Krausz T and Oliva E (2020). Uterine Tumor Resembling Ovarian Sex Cord Stromal Tumor (UTROSCT): A Series of 3 Cases With Extensive Rhabdoid Differentiation, Malignant Behavior, and ESR1-NCOA2 Fusions. *Am J Surg Pathol* 44(11):1563-1572.  41 Goebel EA, Hernandez Bonilla S, Dong F, Dickson BC, Hoang LN, Hardisson D, Lacambra MD, Lu FI, Fletcher CDM, Crum CP, Antonescu CR, Nucci MR and Kolin DL (2020). Uterine Tumor Resembling Ovarian Sex Cord Tumor (UTROSCT): A Morphologic and Molecular Study of 26 Cases Confirms Recurrent NCOA1-3 Rearrangement. *Am J Surg Pathol* 44(1):30-42.  42 Croce S, Lesluyes T, Delespaul L, Bonhomme B, Pérot G, Velasco V, Mayeur L, Rebier F, Ben Rejeb H, Guyon F, McCluggage WG, Floquet A, Querleu D, Chakiba C, Devouassoux-Shisheboran M, Mery E, Arnould L, Averous G, Soubeyran I, Le Guellec S and Chibon F (2019). GREB1-CTNNB1 fusion transcript detected by RNA-sequencing in a uterine tumor resembling ovarian sex cord tumor (UTROSCT): A novel CTNNB1 rearrangement. *Genes Chromosomes Cancer* 58(3):155-163.  43 Moore M and McCluggage WG (2017). Uterine tumour resembling ovarian sex cord tumour: first report of a large series with follow-up. *Histopathology* 71(5):751-759.  44 Folpe AL, Mentzel T, Lehr HA, Fisher C, Balzer BL and Weiss SW (2005). Perivascular epithelioid cell neoplasms of soft tissue and gynecologic origin: a clinicopathologic study of 26 cases and review of the literature. *Am J Surg Pathol* 29(12):1558-1575.  45 Schoolmeester JK, Howitt BE, Hirsch MS, Dal Cin P, Quade BJ and Nucci MR (2014). Perivascular epithelioid cell neoplasm (PEComa) of the gynecologic tract: clinicopathologic and immunohistochemical characterization of 16 cases. *Am J Surg Pathol* 38(2):176-188.  46 Schoolmeester JK, Dao LN, Sukov WR, Wang L, Park KJ, Murali R, Hameed MR and Soslow RA (2015). TFE3 translocation-associated perivascular epithelioid cell neoplasm (PEComa) of the gynecologic tract: morphology, immunophenotype, differential diagnosis. *Am J Surg Pathol* 39(3):394-404.  47 Bennett JA, Braga AC, Pinto A, Van de Vijver K, Cornejo K, Pesci A, Zhang L, Morales-Oyarvide V, Kiyokawa T, Zannoni GF, Carlson J, Slavik T, Tornos C, Antonescu CR and Oliva E (2018). Uterine PEComas: A Morphologic, Immunohistochemical, and Molecular Analysis of 32 Tumors. *Am J Surg Pathol* 42(10):1370-1383.  48 Vang R and Kempson RL (2002). Perivascular epithelioid cell tumor ('PEComa') of the uterus: a subset of HMB-45-positive epithelioid mesenchymal neoplasms with an uncertain relationship to pure smooth muscle tumors. *Am J Surg Pathol* 26(1):1-13.  49 Haimes JD, Stewart CJR, Kudlow BA, Culver BP, Meng B, Koay E, Whitehouse A, Cope N, Lee JC, Ng T, McCluggage WG and Lee CH (2017). Uterine Inflammatory Myofibroblastic Tumors Frequently Harbor ALK Fusions With IGFBP5 and THBS1. *Am J Surg Pathol* 41(6):773-780.  50 Parra-Herran C, Quick CM, Howitt BE, Dal Cin P, Quade BJ and Nucci MR (2015). Inflammatory myofibroblastic tumor of the uterus: clinical and pathologic review of 10 cases including a subset with aggressive clinical course. *Am J Surg Pathol* 39(2):157-168.  51 Bennett JA, Croce S, Pesci A, Niu N, Van de Vijver K, Burks EJ, Burandt E, Zannoni GF, Rabban JT and Oliva E (2020). Inflammatory Myofibroblastic Tumor of the Uterus: An Immunohistochemical Study of 23 Cases. *Am J Surg Pathol* 44(11):1441-1449.  52 Rabban JT, Zaloudek CJ, Shekitka KM and Tavassoli FA (2005). Inflammatory myofibroblastic tumor of the uterus: a clinicopathologic study of 6 cases emphasizing distinction from aggressive mesenchymal tumors. *Am J Surg Pathol* 29(10):1348-1355.  53 Devereaux KA, Fitzpatrick MB, Hartinger S, Jones C, Kunder CA and Longacre TA (2020). Pregnancy-associated Inflammatory Myofibroblastic Tumors of the Uterus Are Clinically Distinct and Highly Enriched for TIMP3-ALK and THBS1-ALK Fusions. *Am J Surg Pathol* 44(7):970-981.  54 Makhdoum S, Nardi V, Devereaux KA, Kunder CA, Nielsen GP, Oliva E, Young RH and Roberts DJ (2020). Inflammatory myofibroblastic tumors associated with the placenta: a series of 9 cases. *Hum Pathol* 106:62-73.  55 Fang H, Langstraat CL, Visscher DW, Folpe AL and Schoolmeester JK (2018). Epithelioid Inflammatory Myofibroblastic Sarcoma of the Ovary With RANB2-ALK Fusion: Report of a Case. *Int J Gynecol Pathol* 37(5):468-472.  56 Mariño-Enríquez A, Wang WL, Roy A, Lopez-Terrada D, Lazar AJ, Fletcher CD, Coffin CM and Hornick JL (2011). Epithelioid inflammatory myofibroblastic sarcoma: An aggressive intra-abdominal variant of inflammatory myofibroblastic tumor with nuclear membrane or perinuclear ALK. *Am J Surg Pathol* 35(1):135-144.  57 Gleason BC and Hornick JL (2008). Inflammatory myofibroblastic tumours: where are we now? *J Clin Pathol* 61(4):428-437.  58 Ferguson SE, Gerald W, Barakat RR, Chi DS and Soslow RA (2007). Clinicopathologic features of rhabdomyosarcoma of gynecologic origin in adults. *Am J Surg Pathol* 31(3):382-389.  59 McCluggage WG, Lioe TF, McClelland HR and Lamki H (2002). Rhabdomyosarcoma of the uterus: report of two cases, including one of the spindle cell variant. *Int J Gynecol Cancer* 12(1):128-132.  60 Li RF, Gupta M, McCluggage WG and Ronnett BM (2013). Embryonal rhabdomyosarcoma (botryoid type) of the uterine corpus and cervix in adult women: report of a case series and review of the literature. *Am J Surg Pathol* 37(3):344-355.  61 Ordi J, Stamatakos MD and Tavassoli FA (1997). Pure pleomorphic rhabdomyosarcomas of the uterus. *Int J Gynecol Pathol* 16(4):369-377.  62 Schoolmeester JK, Carlson J, Keeney GL, Fritchie KJ, Oliva E, Young RH and Nucci MR (2017). Alveolar Soft Part Sarcoma of the Female Genital Tract: A Morphologic, Immunohistochemical, and Molecular Cytogenetic Study of 10 Cases With Emphasis on its Distinction From Morphologic Mimics. *Am J Surg Pathol* 41(5):622-632.  63 Nielsen GP, Oliva E, Young RH, Rosenberg AE, Dickersin GR and Scully RE (1995). Alveolar soft-part sarcoma of the female genital tract: a report of nine cases and review of the literature. *Int J Gynecol Pathol* 14(4):283-292.  64 Kasashima S, Minato H, Kobayashi M, Ueda Y, Oda Y, Hashimoto S and Inoue M (2007). Alveolar soft part sarcoma of the endometrium with expression of CD10 and hormone receptors. *Apmis* 115(7):861-865.  65 Radig K, Buhtz P and Roessner A (1998). Alveolar soft part sarcoma of the uterine corpus. Report of two cases and review of the literature. *Pathol Res Pract* 194(1):59-63.  66 McDonald AG, Dal Cin P, Ganguly A, Campbell S, Imai Y, Rosenberg AE and Oliva E (2011). Liposarcoma arising in uterine lipoleiomyoma: a report of 3 cases and review of the literature. *Am J Surg Pathol* 35(2):221-227.  67 Roma AA, Allende D, Fadare O, Forscher C and Rutgers JK (2017). On Uterine Angiosarcomas: 2 Additional Cases. *Int J Gynecol Pathol* 36(4):369-371.  68 Kruse AJ, Sep S, Slangen BF, Vandevijver NM, Van Gorp T, Kruitwagen RF and Van de Vijver KK (2014). Angiosarcomas of primary gynecologic origin: a clinicopathologic review and quantitative analysis of survival. *Int J Gynecol Cancer* 24(1):4-12.  69 Schammel DP and Tavassoli FA (1998). Uterine angiosarcomas: a morphologic and immunohistochemical study of four cases. *Am J Surg Pathol* 22(2):246-250.  70 Rabban JT, Devine WP, Sangoi AR, Poder L, Alvarez E, Davis JL, Rudzinski E, Garg K and Bean GR (2020). NTRK fusion cervical sarcoma: a report of three cases, emphasising morphological and immunohistochemical distinction from other uterine sarcomas, including adenosarcoma. *Histopathology* 77(1):100-111.  71 Chiang S, Cotzia P, Hyman DM, Drilon A, Tap WD, Zhang L, Hechtman JF, Frosina D, Jungbluth AA, Murali R, Park KJ, Soslow RA, Oliva E, Iafrate AJ, Benayed R, Ladanyi M and Antonescu CR (2018). NTRK Fusions Define a Novel Uterine Sarcoma Subtype With Features of Fibrosarcoma. *Am J Surg Pathol* 42(6):791-798.  72 Croce S, Hostein I, Longacre TA, Mills AM, Pérot G, Devouassoux-Shisheboran M, Velasco V, Floquet A, Guyon F, Chakiba C, Querleu D, Khalifa E, Mayeur L, Rebier F, Leguellec S, Soubeyran I and McCluggage WG (2019). Uterine and vaginal sarcomas resembling fibrosarcoma: a clinicopathological and molecular analysis of 13 cases showing common NTRK-rearrangements and the description of a COL1A1-PDGFB fusion novel to uterine neoplasms. *Mod Pathol* 32(7):1008-1022.  73 Hodgson A, Pun C, Djordjevic B and Turashvili G (2021). NTRK-rearranged Cervical Sarcoma: Expanding the Clinicopathologic Spectrum. *Int J Gynecol Pathol* 40(1):73-77.  74 Demetri GD, Antonescu CR, Bjerkehagen B, Bovée J, Boye K, Chacón M, Dei Tos AP, Desai J, Fletcher JA, Gelderblom H, George S, Gronchi A, Haas RL, Hindi N, Hohenberger P, Joensuu H, Jones RL, Judson I, Kang YK, Kawai A, Lazar AJ, Le Cesne A, Maestro R, Maki RG, Martín J, Patel S, Penault-Llorca F, Premanand Raut C, Rutkowski P, Safwat A, Sbaraglia M, Schaefer IM, Shen L, Serrano C, Schöffski P, Stacchiotti S, Sundby Hall K, Tap WD, Thomas DM, Trent J, Valverde C, van der Graaf WTA, von Mehren M, Wagner A, Wardelmann E, Naito Y, Zalcberg J and Blay JY (2020). Diagnosis and management of tropomyosin receptor kinase (TRK) fusion sarcomas: expert recommendations from the World Sarcoma Network. *Ann Oncol* 31(11):1506-1517.  75 Antonescu CR, Dickson BC, Swanson D, Zhang L, Sung YS, Kao YC, Chang WC, Ran L, Pappo A, Bahrami A, Chi P and Fletcher CD (2019). Spindle Cell Tumors With RET Gene Fusions Exhibit a Morphologic Spectrum Akin to Tumors With NTRK Gene Fusions. *Am J Surg Pathol* 43(10):1384-1391.  76 Weisman PS, Altinok M, Carballo EV, Kushner DM, Kram JJF, Ladanyi M, Chiang S, Buehler D and Dickson Michelson EL (2020). Uterine Cervical Sarcoma With a Novel RET-SPECC1L Fusion in an Adult: A Case Which Expands the Homology Between RET-rearranged and NTRK-rearranged Tumors. *Am J Surg Pathol* 44(4):567-570.  77 Kao YC, Bennett JA, Suurmeijer AJH, Dickson BC, Swanson D, Wanjari P, Zhang L, Lee JC and Antonescu CR (2021). Recurrent MEIS1-NCOA2/1 fusions in a subset of low-grade spindle cell sarcomas frequently involving the genitourinary and gynecologic tracts. *Mod Pathol* 34(6): 1203-1212.  78 Grindstaff SL, DiSilvestro J, Hansen K, DiSilvestro P, Sung CJ and Quddus MR (2020). COL1A1-PDGFB fusion uterine fibrosarcoma: A case report with treatment implication. *Gynecol Oncol Rep* 31:100523.  79 Kolin DL, Quick CM, Dong F, Fletcher CDM, Stewart CJR, Soma A, Hornick JL, Nucci MR and Howitt BE (2020). SMARCA4-deficient Uterine Sarcoma and Undifferentiated Endometrial Carcinoma Are Distinct Clinicopathologic Entities. *Am J Surg Pathol* 44(2):263-270.  80 Kolin DL, Dong F, Baltay M, Lindeman N, MacConaill L, Nucci MR, Crum CP and Howitt BE (2018). SMARCA4-deficient undifferentiated uterine sarcoma (malignant rhabdoid tumor of the uterus): a clinicopathologic entity distinct from undifferentiated carcinoma. *Mod Pathol* 31(9):1442-1456.  81 Lin DI, Allen JM, Hecht JL, Killian JK, Ngo NT, Edgerly C, Severson EA, Ali SM, Erlich RL, Ramkissoon SH, Vergilio JA, Ross JS and Elvin JA (2019). SMARCA4 inactivation defines a subset of undifferentiated uterine sarcomas with rhabdoid and small cell features and germline mutation association. *Mod Pathol* 32(11):1675-1687. | Value list based on the WHO Classification of Female Genital Tumours (2020).  Note that permission to publish the WHO Classification of Tumours may be needed in your implementation. It is advisable to check with the International Agency for Research on Cancer (IARC). |
| Core | MITOTIC COUNTa | \_\_\_\_ /mm2   * Cannot be assessed | The clinical significance of mitotic activity depends on the specific tumour-type involved. Documentation of mitotic activity (highest mitotic count) is required for leiomyosarcoma, STUMP and PEComa, and strongly recommended for UTROSCT, IMT, solitary fibrous tumour and undifferentiated uterine sarcoma. It is optional for other sarcoma types. The 5th edition of the World Health Organization (WHO) Classification of Tumours1 considers both high power fields (HPF) and mm2 for counting of mitoses. In addition, the size of the objective field is mentioned.  For leiomyosarcoma and STUMP, mitotic activity constitutes part of the diagnostic definition together with other histologic features including nuclear atypia and tumour cell necrosis. Mitotic count ≥10 mitoses per 2 mm2 (≥10 mitoses per 10 HPFs if field diameter is 0.55 mm) is used for spindle cell smooth muscle tumours, whereas mitotic count ≥4 mitoses per 2 mm2 (≥4 mitoses per 10 HPFs if field diameter is 0.55 mm) and ≥2 mitoses per 2 mm2 (≥2 mitoses per 10 HPFs if field diameter is 0.55 mm) are used for epithelioid and myxoid smooth muscle tumours, respectively.1 For STUMP, mitotic activity forms part of the diagnostic definition under two scenarios based on the 2020 WHO Classification:1 1) tumours with focal/multifocal or diffuse nuclear atypia, and 5-9 mitoses per 2 mm2 (5-9 mitoses per 10 HPFs if field diameter is 0.55 mm) but lacking tumour cell necrosis; and 2) tumours showing ≥15 mitoses per 2 mm2 (≥15 mitoses per 10 HPFs if field diameter is 0.55 mm) and lacking nuclear atypia and tumour cell necrosis. It is important to note that degenerative nuclear changes/karyorrhexis may mimic mitotic figures, particularly atypical mitotic figures. It is generally recommended that a formal mitotic count should rely predominantly if not exclusively on counting of typical bipolar mitoses. For PEComa, the presence of any mitotic activity, together with tumour size (≥50 mm), high grade atypia, necrosis and lymphovascular invasion form the criteria for malignancy in the gynaecologic tract.2-4 For other rare uterine sarcoma types in which mitotic count is part of the risk stratification (e.g., solitary fibrous tumour), mitotic activity should also be documented.    For IMT, there is limited evidence that mitotic count and large tumour size may be associated with more aggressive clinical behaviour.5,6 For UTROSCT, there is also limited evidence that elevated mitotic counts and necrosis are associated with malignant behaviour.7 Mitotic activity is generally brisk for undifferentiated uterine sarcoma and mitotic count has been shown to be prognostically relevant in undifferentiated uterine sarcomas (lacking endometrial stromal sarcoma genetic fusions) with tumours showing a mitotic count of >25 mitoses per 2 mm2 (>25 mitoses per 10 HPFs if field diameter is 0.55 mm) being associated with decreased survival.8,9  For adenosarcoma, most tumours demonstrate stromal mitoses (>1 mitosis per 2 mm2 (>1 mitosis per 10 HPFs if field diameter is 0.55 mm)) but mitotic activity may be minimal or even absent in some cases.10,11 There is currently no evidence that mitotic count alone is prognostically significant, in contrast to the presence of sarcomatous overgrowth and/or deep myometrial invasion which are associated with worse prognosis.12-14 With regard to endometrial stromal sarcomas, while low grade endometrial stromal sarcomas tend to exhibit lower mitotic counts than high grade endometrial stromal sarcomas, there is overlap in the range of mitotic activity and the number of mitoses is not used for diagnostic classification. However, most low grade endometrial stromal sarcomas display a mitotic rate of <5 mitoses per 2 mm2 (<5 mitoses per 10 HPF if field diameter is 0.55 mm) and a finding of high mitotic rate (particularly >10 mitoses) should prompt more thorough tumour sampling and careful histologic evaluation as well as consideration of ancillary studies to exclude high grade endometrial stromal sarcoma or other tumour types. The degree of mitotic activity has no diagnostic or known prognostic significance for recently recognised entities including *SMARCA4*-deficient uterine sarcoma and *NTRK*-rearranged sarcoma. Mitotic activity is typically high in *SMARCA4*-deficient uterine sarcoma and is variable in *NTRK*-rearranged sarcoma.  **References**  1 WHO Classification of Tumours Editorial Board (2020). *Female Genital Tumours, WHO Classification of Tumours, 5th Edition, Volume 4*. IARC Press, Lyon.  2 Bennett JA, Braga AC, Pinto A, Van de Vijver K, Cornejo K, Pesci A, Zhang L, Morales-Oyarvide V, Kiyokawa T, Zannoni GF, Carlson J, Slavik T, Tornos C, Antonescu CR and Oliva E (2018). Uterine PEComas: A Morphologic, Immunohistochemical, and Molecular Analysis of 32 Tumors. *Am J Surg Pathol* 42(10):1370-1383.  3 Fadare O (2008). Perivascular epithelioid cell tumor (PEComa) of the uterus: an outcome-based clinicopathologic analysis of 41 reported cases. *Adv Anat Pathol* 15(2):63-75.  4 Folpe AL, Mentzel T, Lehr HA, Fisher C, Balzer BL and Weiss SW (2005). Perivascular epithelioid cell neoplasms of soft tissue and gynecologic origin: a clinicopathologic study of 26 cases and review of the literature. *Am J Surg Pathol* 29(12):1558-1575.  5 Parra-Herran C, Schoolmeester JK, Yuan L, Dal Cin P, Fletcher CD, Quade BJ and Nucci MR (2016). Myxoid Leiomyosarcoma of the Uterus: A Clinicopathologic Analysis of 30 Cases and Review of the Literature With Reappraisal of Its Distinction From Other Uterine Myxoid Mesenchymal Neoplasms. *Am J Surg Pathol* 40(3):285-301.  6 Bennett JA, Nardi V, Rouzbahman M, Morales-Oyarvide V, Nielsen GP and Oliva E (2017). Inflammatory myofibroblastic tumor of the uterus: a clinicopathological, immunohistochemical, and molecular analysis of 13 cases highlighting their broad morphologic spectrum. *Mod Pathol* 30(10):1489-1503.  7 Moore M and McCluggage WG (2017). Uterine tumour resembling ovarian sex cord tumour: first report of a large series with follow-up. *Histopathology* 71(5):751-759.  8 Gremel G, Liew M, Hamzei F, Hardell E, Selling J, Ghaderi M, Stemme S, Pontén F and Carlson JW (2015). A prognosis based classification of undifferentiated uterine sarcomas: identification of mitotic index, hormone receptors and YWHAE-FAM22 translocation status as predictors of survival. *Int J Cancer* 136(7):1608-1618.  9 Hardell E, Josefson S, Ghaderi M, Skeie-Jensen T, Westbom-Fremer S, Cheek EH, Bell D, Selling J, Schoolmeester JK, Måsbäck A, Davidson B and Carlson JW (2017). Validation of a Mitotic Index Cutoff as a Prognostic Marker in Undifferentiated Uterine Sarcomas. *Am J Surg Pathol* 41(9):1231-1237.  10 Howitt BE et al (eds) (2020). Adenosarcoma of the uterine corpus. In: *Female Genital Tumours, WHO Classification of Tumours, 5th Edition, Volume 4*, WHO Classification of Tumours Editorial Board (ed), IARC Press, Lyon.  11 Yemelyanova A et al (eds) (2020). Adenosarcoma of the uterine cervix. In: *Female Genital Tumours, WHO Classification of Tumours, 5th Edition, Volume 4*, WHO Classification of Tumours Editorial Board (ed), IARC Press, Lyon.  12 Clement PB and Scully RE (1990). Mullerian adenosarcoma of the uterus: a clinicopathologic analysis of 100 cases with a review of the literature. *Hum Pathol* 21(4):363-381.  13 Clement PB (1989). Müllerian adenosarcomas of the uterus with sarcomatous overgrowth. A clinicopathological analysis of 10 cases. *Am J Surg Pathol* 13(1):28-38.  14 Zaloudek CJ and Norris HJ (1981). Adenofibroma and adenosarcoma of the uterus: a clinicopathologic study of 35 cases. *Cancer* 48(2):354-366. | a Core for leiomyosarcoma, STUMP, PEComa; non-core for all  other entities but including mitotic count is strongly  recommended. |
| Core and Non-core | **EXTENT OF INVASION** | **Myometrial or cervical stromal invasion**  *(Applicable to adenosarcoma only)*   * Cannot be assessed * Not identified * ≤50% * >50%   **Uterine serosa involvement**   * Cannot be assessed * Not involved   Distance of tumour to uterine serosa \_\_\_ mm   * Involved   **Parametrial involvement**   * Not submitted * Cannot be assessed * Not involved * Involved * Left * Right * Indetermine   **Omentumb**   * Cannot be assessed * Not involved * Involved   Vaginab   * Cannot be assessed * Not involved * Involved   Fallopian tubeb   * Cannot be assessed * Not involved * Involved * Left * Right * Indetermine   Ovaryb   * Cannot be assessed * Not involved * Involved * Left * Right * Indetermine   Peritoneal biopsiesb   * Not involved * Involved   Peritoneal washings/peritoneal fluidb   * Positive * Negative * Atypical/suspicious | Myometrial or cervical stromal invasion  The depth of myometrial invasion is an essential parameter in the staging of adenosarcomas located in the uterine corpus or cervix (see **PROVISIONAL PATHOLOGICAL STAGING**). According to the current International Federation of Gynecology and Obstetrics (FIGO) Staging System,1 Stage IA adenosarcoma is limited to the endometrium/endocervix; Stage IB invades ≤50% of the myometrium or cervical stroma; and Stage IC invades more than 50% of the myometrium or cervical stroma.2,3 Myometrial infiltration is also an important prognostic factor in uterine adenosarcoma for overall survival and recurrence.4-6  Because the staging of low and high grade endometrial stromal sarcomas, leiomyosarcoma and undifferentiated uterine sarcoma (and other sarcomas) is not based currently on myometrial infiltration, the depth of myometrial infiltration is not relevant.  Uterine serosa involvement  Uterine serosal involvement should be documented as it is an adverse prognostic factor in uterine leiomyosarcoma.7 Although evidence for clinical relevance for other uterine sarcomas is limited, ICCR Uterine Sarcoma Dataset Authoring Committee (DAC) considers it to represent a core element in reporting.  Tumour-free distance to uterine serosa refers to the distance between the deepest point of tumour within the myometrium and the nearest serosal surface and is considered a non-core element.  Parametrial involvement  Parametrium is defined as the fibro-adipose connective tissue located laterally in the supracervical portion of the uterus. Most hysterectomies for uterine sarcoma will be simple hysterectomies without parametrial resections. If parametrial tissue is removed, the presence or absence of parametrial involvement should be documented as best practice. Although evidence for clinical relevance of parametrial involvement in uterine sarcomas is limited, the DAC considers it to represent a core element in reporting.  Omentum  Omental involvement should be documented as it contributes to the staging assessment. FIGO Stage IIIA equates to one site of abdominal involvement and IIIB to more than one site.1  Vagina  A total hysterectomy can have a vaginal cuff which should be measured. The presence or absence of vaginal involvement in such cases should be documented on the report.  Fallopian tube  The presence or absence of adnexal (ovarian/fallopian tube) involvement should be documented. Adnexal involvement affects the tumour stage (FIGO Stage IIA) which remains the most powerful prognostic factor for uterine sarcomas,3,5,8,9 and may occur as a result of direct extension or metastatic spread of tumour.  Ovary  The presence or absence of adnexal (ovarian/fallopian tube) involvement should be documented. Adnexal involvement affects the tumour stage (FIGO Stage IIA) which remains the most powerful prognostic factor for uterine sarcomas,3,5,8,9 and may occur as a result of direct extension or metastatic spread of tumour.  Peritoneal biopsies  Peritoneal involvement should be documented as it contributes to the staging assessment. Pelvic peritoneal involvement equates to FIGO Stage IIB while abdominal peritoneal involvement equates to FIGO Stage IIIA or IIIB depending on the number of sites involved.1  Peritoneal washings/peritoneal fluid  The presence or absence of tumour cells in peritoneal fluid/washings should be documented if this specimen type is submitted. There is only limited data suggesting that positive peritoneal cytology may be an adverse prognostic factor in uterine sarcomas with one study suggesting that positive peritoneal cytology may be a prognostic factor for mortality in uterine sarcomas, particularly in leiomyosarcoma.10 Accrual of this data prospectively will facilitate future study regarding the prognostic significance of positive peritoneal fluid.  **References**  1 Prat J and Mbatani (2015). Uterine sarcomas. *Int J Gynaecol Obstet* 131 Suppl 2:S105-110.  2 Prat J (2009). FIGO staging for uterine sarcomas. *Int J Gynaecol Obstet* 104(3):177-178.  3 Mbatani N, Olawaiye AB and Prat J (2018). Uterine sarcomas. *Int J Gynaecol Obstet* 143 Suppl 2:51-58.  4 Hodgson A, Amemiya Y, Seth A, Djordjevic B and Parra-Herran C (2017). High-grade Müllerian Adenosarcoma: Genomic and Clinicopathologic Characterization of a Distinct Neoplasm With Prevalent TP53 Pathway Alterations and Aggressive Behavior. *Am J Surg Pathol* 41(11):1513-1522.  5 Abeler VM, Røyne O, Thoresen S, Danielsen HE, Nesland JM and Kristensen GB (2009). Uterine sarcomas in Norway. A histopathological and prognostic survey of a total population from 1970 to 2000 including 419 patients. *Histopathology* 54(3):355-364.  6 Gallardo A and Prat J (2009). Mullerian adenosarcoma: a clinicopathologic and immunohistochemical study of 55 cases challenging the existence of adenofibroma. *Am J Surg Pathol* 33(2):278-288.  7 Tirumani SH, Deaver P, Shinagare AB, Tirumani H, Hornick JL, George S and Ramaiya NH (2014). Metastatic pattern of uterine leiomyosarcoma: retrospective analysis of the predictors and outcome in 113 patients. *J Gynecol Oncol* 25(4):306-312.  8 Amant F, Coosemans A, Debiec-Rychter M, Timmerman D and Vergote I (2009). Clinical management of uterine sarcomas. *Lancet Oncol* 10(12):1188-1198.  9 Ricci S, Stone RL and Fader AN (2017). Uterine leiomyosarcoma: Epidemiology, contemporary treatment strategies and the impact of uterine morcellation. *Gynecol Oncol* 145(1):208-216.  10 Matsuo K, Matsuzaki S, Nusbaum DJ, Ki S, Chang EJ, Klar M and Roman LD (2021). Significance of Malignant Peritoneal Cytology on Survival of Women with Uterine Sarcoma. *Ann Surg Oncol* 28(3):1740-1748. | b If received. |
| Core | LYMPHOVASCULAR INVASION | * Indeterminate * Not identified * Present | The presence or absence of lymphovascular invasion (LVI) in uterine sarcomas should be documented. For some tumours, such as low grade and high grade endometrial stromal sarcoma, LVI is frequently encountered; in contrast, other tumours, such as adenosarcomas, uncommonly demonstrate LVI unless associated with deep myometrial invasion and/or sarcomatous overgrowth. The presence of LVI may carry prognostic significance in leiomyosarcoma, particularly when early stage,1,2 and in adenosarcoma.3-5  One study evaluated specific patterns of vascular involvement by low grade endometrial stromal sarcoma, high grade endometrial stromal sarcoma, leiomyosarcoma, and undifferentiated uterine sarcoma and divided patterns into ‘true’ vascular invasion versus ‘intrusion’ into lymphovascular spaces.6 True LVI was characterised by dyscohesive clusters of tumours cells with irregular edges, lacking vasculature within the intravascular tumour and/or lack of immunohistochemically proven endothelial cells surrounding the intravascular tumour focus. Vascular intrusion, which was considered to be ‘pseudoinvasion’, was characterised by cohesive intravascular tumour with smooth contours and lined by endothelial cells. Pre-existing vascular spaces were frequently identified within the intravascular tumour in the cases of vascular intrusion and such foci were often in direct communication with the main tumour mass.6  While usually straightforward, the assessment of LVI may be difficult in a minority of cases, for which the reasons may include (but are not limited to) suboptimal fixation or cauterization artefacts. In such cases, examination of multiple levels and/or immunostaining for endothelial or lymphatic markers (such as CD31, CD34, D2-40 and ERG) may be employed to assist with the decision-making. Cases that are still equivocal after taking additional steps may be reported as ‘indeterminate’ for LVI, but this designation should only be sparingly used and it is useful to provide the reason in a comment in the report.  **References**  1 Vaz J, Tian C, Richardson MT, Chan JK, Mysona D, Rao UN, Powell MA, Shriver CD, Hamilton CA, Casablanca Y, Maxwell GL and Darcy KM (2020). Impact of adjuvant treatment and prognostic factors in stage I uterine leiomyosarcoma patients treated in Commission on Cancer®-accredited facilities. *Gynecol Oncol* 157(1):121-130.  2 Singh N, Al-Ruwaisan M, Batra A, Itani D and Ghatage P (2020). Factors Affecting Overall Survival in Premenopausal Women With Uterine Leiomyosarcoma: A Retrospective Analysis With Long-Term Follow-Up. *J Obstet Gynaecol Can* 42(12):1483-1488.  3 Nathenson MJ, Conley AP, Lin H, Fleming N, Lazar A, Wang WL and Ravi V (2018). The Importance of Lymphovascular Invasion in Uterine Adenosarcomas: Analysis of Clinical, Prognostic, and Treatment Outcomes. *Int J Gynecol Cancer* 28(7):1297-1310.  4 Carroll A, Ramirez PT, Westin SN, Soliman PT, Munsell MF, Nick AM, Schmeler KM, Klopp AH and Fleming ND (2014). Uterine adenosarcoma: an analysis on management, outcomes, and risk factors for recurrence. *Gynecol Oncol* 135(3):455-461.  5 Yuan Z, Shen K, Yang J, Cao D, Zhang Y, Zhou H, Wu H and Yu M (2019). Uterine Adenosarcoma: A Retrospective 12-Year Single-Center Study. *Front Oncol* 9:237.  6 Roma AA, Barbuto DA, Samimi SA, Stolnicu S, Alvarado-Cabrero I, Chanona-Vilchis J, Aguilera-Barrantes I, de Peralta-Venturina M, Malpica A, Rutgers JK and Silva EG (2015). Vascular invasion in uterine sarcomas and its significance. A multi-institutional study. *Hum Pathol* 46(11):1712-1721. |  |
| Core and Non-core | MARGIN STATUS | Distal/cervical or vaginal   * Cannot be assessed * Not involved   Distance of tumour from closest  margin \_\_\_ mm  Specify closest margin, if  possible   * Involved   Specify margin, if possible   * Cervical * Vaginal * Other, *specify*   Parametrial   * Cannot be assessed * Not involved   Specify laterality, if possible   * Involved | Margins of resection are an important parameter to include in reporting as they may guide post-surgical treatment with chemotherapy and/or radiation therapy depending on tumour type. In addition, positive margins have been shown to be a negative prognostic factor for a variety of different uterine sarcomas including low and high grade endometrial stromal sarcoma,1 leiomyosarcoma,2 and Müllerian adenosarcoma.3 The most relevant margin is usually the distal cervicovaginal resection margin. However, the status of other surgical margins, such as the parametrium (when removed) including laterality, should also be documented.  **References**  1 Seagle BL, Shilpi A, Buchanan S, Goodman C and Shahabi S (2017). Low-grade and high-grade endometrial stromal sarcoma: A National Cancer Database study. *Gynecol Oncol* 146(2):254-262.  2 Seagle BL, Sobecki-Rausch J, Strohl AE, Shilpi A, Grace A and Shahabi S (2017). Prognosis and treatment of uterine leiomyosarcoma: A National Cancer Database study. *Gynecol Oncol* 145(1):61-70.  3 Seagle BL, Kanis M, Strohl AE and Shahabi S (2016). Survival of women with Mullerian adenosarcoma: A National Cancer Data Base study. *Gynecol Oncol* 143(3):636-641. |  |
| Core | LYMPH NODE STATUSc | Pelvic nodes   * Cannot be assessed * No nodes submitted or found   Number of nodes examined  Number of positive nodes  Size of maximum tumour  deposit \_\_\_ mm  Para-aortic nodes   * Cannot be assessed * No nodes submitted or found   Number of nodes examined  Number of positive nodes  Size of maximum tumour  deposit \_\_\_ mm  **Other lymph nodes removed, *specify site(s) \_\_\_\_\_\_\_\_\_\_\_\_***  Number of nodes examined  Number of positive nodes  Size of maximum tumour  deposit \_\_\_ mm | The anatomic location and number of lymph nodes dissected, the number containing tumour and the size of the largest tumour deposit should be accurately documented in the pathology report. According to TNM8,1 nodal involvement should be recorded as the presence of isolated tumour cells (ITC, <0.2 mm), micrometastases (MIC, 0.2-2 mm) or macrometastases (MAC, >2 mm). MAC are regarded as pN1, MIC as pN1 (mi) and ITCs are pN0 (i+); ITCs do not upstage a neoplasm. Involvement of pelvic and/or para-aortic lymph nodes by uterine sarcoma will upstage the sarcoma. The number of lymph nodes examined and number of lymph nodes involved by tumour should be reported for regional lymphadenectomies, if performed.    Because of the low risk of metastatic disease in lymph nodes, routine lymph node dissection is typically not undertaken in low stage uterine leiomyosarcomas.2-4 Moreover, lymphadenectomy is not routinely undertaken for uterine leiomyosarcoma as it does not appear to impact overall survival.4-6 Nevertheless, lymph node resection should be performed if the lymph nodes appears enlarged or suspicious.7 The reported frequency of lymph node involvement in low grade endometrial stromal sarcomas ranges from 3.6% to 10%,8-10 and from 10.2% to 44% for high grade endometrial stromal sarcoma.8,11 The prognostic importance of lymphadenectomy for endometrial stromal sarcomas has been a subject of debate,9,12-15 although a recent meta-analysis concluded that for localised endometrial stromal sarcoma and leiomyosarcoma, lymphadenectomy is not recommended.6  Lymph node metastasis is a significant prognostic factor in uterine adenosarcoma.16 However, lymphadenectomy is not typically performed unless lymph nodes appear enlarged and/or suspicious as the rate of nodal metastasis is low (6.5%).17  **References**  1 Brierley JD, Gospodarowicz MK and Wittekind C (eds) (2016). *Union for International Cancer Control. TNM Classification of Malignant Tumours, 8th Edition*, Wiley, USA.  2 George S, Serrano C, Hensley ML and Ray-Coquard I (2018). Soft Tissue and Uterine Leiomyosarcoma. *J Clin Oncol* 36(2):144-150.  3 Goff BA, Rice LW, Fleischhacker D, Muntz HG, Falkenberry SS, Nikrui N and Fuller AF, Jr. (1993). Uterine leiomyosarcoma and endometrial stromal sarcoma: lymph node metastases and sites of recurrence. *Gynecol Oncol* 50(1):105-109.  4 Nesrine T, Ines Z, Abdelwahed N, Ali AM, Nadia B, Monia H, Hatem B, Maher S and Khaled R (2019). Prognostic factors and the role of pelvic lymphadenectomy in uterine leiomyosarcomas. *SAGE Open Med* 7:2050312119856817.  5 Seagle BL, Sobecki-Rausch J, Strohl AE, Shilpi A, Grace A and Shahabi S (2017). Prognosis and treatment of uterine leiomyosarcoma: A National Cancer Database study. *Gynecol Oncol* 145(1):61-70.  6 Si M, Jia L, Song K, Zhang Q and Kong B (2017). Role of Lymphadenectomy for Uterine Sarcoma: A Meta-Analysis. *Int J Gynecol Cancer* 27(1):109-116.  7 Leitao MM, Sonoda Y, Brennan MF, Barakat RR and Chi DS (2003). Incidence of lymph node and ovarian metastases in leiomyosarcoma of the uterus. *Gynecol Oncol* 91(1):209-212.  8 Seagle BL, Shilpi A, Buchanan S, Goodman C and Shahabi S (2017). Low-grade and high-grade endometrial stromal sarcoma: A National Cancer Database study. *Gynecol Oncol* 146(2):254-262.  9 Zhang Y, Li N, Wang W, Yao H, An J, Li N, Sun Y and Wu L (2020). Long-term impact of lymphadenectomies in patients with low-grade, early-stage uterine endometrial stroma sarcoma. *J Obstet Gynaecol Res* 46(4):654-662.  10 Dos Santos LA, Garg K, Diaz JP, Soslow RA, Hensley ML, Alektiar KM, Barakat RR and Leitao MM, Jr. (2011). Incidence of lymph node and adnexal metastasis in endometrial stromal sarcoma. *Gynecol Oncol* 121(2):319-322.  11 Malouf GG, Lhommé C, Duvillard P, Morice P, Haie-Meder C and Pautier P (2013). Prognostic factors and outcome of undifferentiated endometrial sarcoma treated by multimodal therapy. *Int J Gynaecol Obstet* 122(1):57-61.  12 Amant F, Coosemans A, Debiec-Rychter M, Timmerman D and Vergote I (2009). Clinical management of uterine sarcomas. *Lancet Oncol* 10(12):1188-1198.  13 Barney B, Tward JD, Skidmore T and Gaffney DK (2009). Does radiotherapy or lymphadenectomy improve survival in endometrial stromal sarcoma? *Int J Gynecol Cancer* 19(7):1232-1238.  14 Zhou J, Zheng H, Wu SG, He ZY, Li FY, Su GQ and Sun JY (2015). Influence of different treatment modalities on survival of patients with low-grade endometrial stromal sarcoma: A retrospective cohort study. *Int J Surg* 23(Pt A):147-151.  15 Shah JP, Bryant CS, Kumar S, Ali-Fehmi R, Malone JM, Jr. and Morris RT (2008). Lymphadenectomy and ovarian preservation in low-grade endometrial stromal sarcoma. *Obstet Gynecol* 112(5):1102-1108.  16 Nathenson MJ and Conley AP (2018). Prognostic factors for uterine adenosarcoma: a review. *Expert Rev Anticancer Ther* 18(11):1093-1100.  17 Tropé CG, Abeler VM and Kristensen GB (2012). Diagnosis and treatment of sarcoma of the uterus. A review. *Acta Oncol* 51(6):694-705. | c If resected. |
| Non-core | COEXISTENT PATHOLOGY | * None identified * Present, *specify* | There are no known precursor lesions of uterine sarcomas. Unrelated incidental conditions can be documented. |  |
| Non-core | ANCILLARY STUDIES | * Not performed * Performed * Immunohistochemistry, *specify test(s) and result(s)* * Molecular findings, *specify test(s) and result(s)* * Other*,* *specify test(s) and result(s)*   **Representative blocks for ancillary studies**, *specify those blocks best representing tumour and/or normal tissue for further study* | Ancillary testing (chiefly immunohistochemistry and/or molecular testing) may be of value in the diagnosis of uterine malignant and potentially malignant mesenchymal tumours. The results of ancillary tests should be interpreted in the overall context of the clinical setting, macroscopic pathology and microscopic pathology. The most recent World Health Organization (WHO) Classification1 defines two potential roles for ancillary tests for certain tumours: 1) to serve as essential diagnostic criteria, required for establishing the diagnosis; or 2) to serve as supportive criteria that are desirable but not essential to establish the diagnosis. To harmonise with the latest WHO Classification,1 the International Collaboration on Cancer Reporting Uterine Sarcoma Dataset Authoring Committee recommends adopting a similar strategy, acknowledging that some of these ancillary tests may not be available in all practice settings. Discussion of the detailed immunophenotype of the various uterine sarcomas or use of ancillary testing to resolve specific differential diagnoses is beyond the scope of these recommendations. Among the tumours with a defining molecular alteration, gene fusion is the main pathologic mechanism; thus, fluorescent in situ hybridisation or RNA sequencing are the primary types of molecular diagnostic tools, with a few rare exceptions of tumours characterised by inactivating mutations.  Leiomyosarcoma and STUMP are expected to exhibit a smooth muscle immunophenotype (positive for desmin, h-caldesmon, smooth muscle myosin and smooth muscle actin), although it is not uncommon for only some of the smooth muscle stains to be positive or for staining to be patchy, particularly in myxoid and epithelioid variants. While mutation of *TP53*, *MED12*,and/or *ATRX* occur in a minority of leiomyosarcomas, these alterations are not specific to leiomyosarcoma. A recent study has shown that p53 immunohistochemistry may be useful in distinguishing translocated associated sarcomas from other non-translocated associated sarcomas with the former more often showing wild-type staining.2 A minority of myxoid leiomyosarcoma may exhibit PLAG1 immunoreactivity and *PLAG1* fusion. Low grade endometrial stromal sarcoma is expected to exhibit diffuse strong CD10 and estrogen receptor (ER) immunoreactivity The diagnosis can be supported by demonstrating a gene fusion involving *JAZF1* and/or *PHF1* but since only about two-thirds of these tumours harbor such a gene fusion, molecular testing is not essential for the diagnosis nor does a negative result exclude the diagnosis. High grade endometrial stromal sarcoma encompasses a range of tumours that are subclassified by one of a variety of distinct gene fusions, thus requiring molecular testing for their diagnosis. The high grade component of *YWHAE-NUTM2A/B* high grade endometrial stromal sarcoma typically exhibits absent CD10 and ER immunoreactivity, positive cyclin D1, CD117, CD56, CD99 and BCOR immunoreactivity,3 and the *YWHAE-NUTM2A/B* gene fusion. *ZC3H7B-BCOR* high grade endometrial stromal sarcoma retains CD10 immunoreactivity, exhibits variable ER immunoexpression, positive cyclin D1 immunoreactivity, variable BCOR immunoreactivity, and *ZC3H7B-BCOR* gene fusion. High grade endometrial stromal sarcoma with *BCOR* internal tandem duplication (ITD) exhibit variable CD10 immunoexpression, loss of ER immunoreactivity, positive cyclin D1 and BCOR immunoreactivity, and *BCOR* ITD by molecular sequencing techniques.  *SMARCA4-*deficient uterine sarcoma is defined by loss of SMARCA4 (BRG1) immunoreactivity or, rarely, loss of SMARCB1 (INI1) immunoreactivity. The diagnosis can be supported by demonstrating inactivating mutation or deletion of *SMARCA4.* IMT is defined by positive ALK immunoreactivity; demonstration of *ALK* fusion by molecular testing can support the diagnosis but is not essential if the ALK immunostain is positive. PEComa is defined by dual melanocytic (HMB45, cathepsin K, melan A, MITF, and/or PNL2) and myoid (smooth muscle actin, desmin, h-caldesmon) immunoreactivity. It is recommended that at least two melanocytic markers be positive given the lack of specificity of any one marker for PEComa. The subset of PEComas that harbour a *TFE3* fusion exhibit TFE3 immunoreactivity along with melanocytic marker immunoreactivity, although smooth muscle marker immunoreactivity may be limited or absent. The diagnosis of PEComa can be supported by demonstration of an inactivating mutation of *TSC1* or *TSC2* or by demonstrating *TFE3* or *RAD51B* fusion.4 UTROSCT is characterised by polyphenotypic immunoreactivity of epithelial markers (keratin, epithelial membrane antigen (EMA)), sex cord markers (FOXL2, SF1, calretinin, inhibin, WT1, and/or melan A), myoid markers (smooth muscle actin, desmin and h-caldesmon) and hormone receptors (ER and progesterone receptor (PR)). The diagnosis can be supported by demonstrating *ESR1* or *GREB1* fusion; however such alterations are not present in all cases, so a negative result does not exclude the diagnosis. *NTRK* uterine sarcoma is defined by a gene fusion involving *NTRK1, NTRK2,* or *NTKR3.* S100 and CD34 are usually positive and immunoreactivity of these markers can be used as a screening tool to identify tumours that merit *NTRK* molecular testing; smooth muscle markers, CD10 and hormone receptors are usually negative. Pan-TRK immunoreactivity can also be used to triage testing for a *NTRK* fusion. However, high grade endometrial stromal sarcoma may show NTRK immunoreactivity in the absence of an *NTRK* fusion.5Uterine adenosarcomas do not have a unique immunophenotype and so the diagnosis is mainly based on morphologic criteria. The tumour cells usually exhibit CD10 and ER immunoreactivity but these markers may be absent in areas of high grade stroma/sarcomatous overgrowth. Rhabdomyosarcoma is expected to exhibit immunoreactivity of desmin, myogenin, and/or myoD1. Both rhabdomyosarcoma and adenosarcoma with rhabdomyosarcomatous differentiation may harbor *DICER1* mutations.  **References**  1 WHO Classification of Tumours Editorial Board (2020). *Female Genital Tumours, WHO Classification of Tumours, 5th Edition, Volume 4*. IARC Press, Lyon.  2 Mohammad N, Stewart CJR, Chiang S, Turashvili G, Dickson BC, Ng TL, Köbel M, McCluggage WG, Croce S and Lee CH (2020). p53 immunohistochemical analysis of fusion-positive uterine sarcomas. *Histopathology*.  3 McCluggage WG and Lee CH (2019). YWHAE-NUTM2A/B Translocated High-grade Endometrial Stromal Sarcoma Commonly Expresses CD56 and CD99. *Int J Gynecol Pathol* 38(6):528-532.  4 Selenica P, Conlon N, Gonzalez C, Frosina D, Jungbluth AA, Beets-Tan RGH, Rao MK, Zhang Y, Benayed R, Ladanyi M, Solit DB, Chiang S, Hyman DM, Hensley ML, Soslow RA, Weigelt B and Murali R (2021). Genomic Profiling Aids Classification of Diagnostically Challenging Uterine Mesenchymal Tumors With Myomelanocytic Differentiation. *Am J Surg Pathol* 45(1):77-92.  5 Momeni-Boroujeni A, Mohammad N, Wolber R, Yip S, Köbel M, Dickson BC, Hensley ML, Leitao MM, Jr., Antonescu CR, Benayed R, Ladanyi M, Lee CH and Chiang S (2021). Targeted RNA expression profiling identifies high-grade endometrial stromal sarcoma as a clinically relevant molecular subtype of uterine sarcoma. *Mod Pathol* 34(5):1008-1016. |  |
| Core | PATHOLOGICALLY CONFIRMED DISTANT METASTASIS | * Not identified * Present, *specify site(s)* | Documentation of known metastatic disease is an important part of the pathology report. Such information, if available, should be recorded with as much detail as is available including the site, whether the specimen is a histopathology or cytopathology specimen and with reference to any relevant prior surgical pathology or cytopathology specimens. |  |

| **Core/**  **Non-core** | **Element name** | **Values** | **Commentary** | **Implementation notes** |
| --- | --- | --- | --- | --- |
| Core | PROVISIONAL PATHOLOGICAL STAGING | **FIGO (2001 edition)d**  Leiomyosarcomas and endometrial stromal sarcomas   * I Tumour limited to uterus * IA Less than 5 cm * IB More than 5 cm * II Tumour extends beyond the uterus, within the pelvis * IIA Adnexal involvement * IIB Involvement of other pelvic tissues * III Tumour invades abdominal tissues * (not just protruding into the abdomen) * IIIA One site * IIIB More than one site * IIIC Metastasis to pelvic and/or para-aortic lymph nodes * IV Tumour invades bladder and/or rectum and/or distant metastasis * IVA Tumour invades bladder and/or rectum * IVB Distant metastasis   Adenosarcomas   * I Tumour limited to uterus * IA Tumour limited to endometrium/endocervix with no myometrial invasion * IB Less than or equal to half myometrial invasion * IC More than half myometrial invasion * II Tumour extends to the pelvis * IIA Adnexal involvement * IIB Tumour extends to extrauterine pelvic tissue * III Tumour invades abdominal tissues (not just protruding into the abdomen) * IIIA One site * IIIB More than one site * IIIC Metastasis to pelvic and/or para-aortic lymph nodes * IV Tumour invades bladder and/or rectum and/or distant metastasis * IVA Tumour invades bladder and/or rectum * IVB Distant metastasis   **TNM Staging (UICC TNM 8th edition 2016)e**  **TNM Descriptors**  (only if applicable)   * m - multiple primary tumours * r - recurrent * y - post-therapy   **Primary tumour (pT)**  LEIOMYOSARCOMAS AND ENDOMETRIAL STROMAL SARCOMASf   * T1 Tumour limited to the uterus * T1a Tumour 5 cm or less in greatest dimension * T1b Tumour more than 5 cm * T2 Tumour extends beyond the uterus, within the pelvis * T2a Tumour involves adnexa * T2b Tumour involves other pelvis tissues * T3 Tumour infiltrates abdominal tissues * T3a One site * T3b More than one site * N1 Metastasis to regional lymph nodes * T4 Tumour invades bladder or rectum * M1 Distant metastasis   ADENOSARCOMA   * T1 Tumour limited to the uterus * T1a Tumour limited to the endometrium/endocervix * T1b Tumour invades to less than half of the myometrium * T1c Tumour invades more than half of the myometrium * T2 Tumour extends beyond the uterus, within the pelvis * T2a Tumour involves adnexa * T2b Tumour involves other pelvis tissues * T3 Tumour involves abdominal tissues * T3a One site * T3b More than one site * N1 Metastasis to regional lymph nodes * T4 Tumour invades bladder or rectum * M1 Distant metastasis   **Regional lymph nodes (pN)**   * NX Regional lymph nodes cannot be assessed * N0 No regional lymph node metastasis * N1 Regional lymph node metastasis | The pathological staging must be provided on the pathology report and is therefore a core element. The term ‘provisional pathological staging’ is used in this dataset to indicate that the stage that is provided may not represent the final tumour stage which should be determined at the multidisciplinary tumour board meeting where all the pathological, clinical and radiological features are available.1-3  The latest version of either International Federation of Gynecology and Obstetrics (FIGO) *or* TNM staging, *or* both, can be used depending on local preferences.1-3 The FIGO Staging System is in widespread use internationally and is the system used in most clinical trials and research studies. However, Union for International Cancer Control (UICC) or American Joint Committee on Cancer (AJCC) versions of TNM are used or mandated in many parts of the world.2,3 With regards to updating of staging systems, there is collaboration between FIGO and those agencies responsible for TNM with an agreement to adopt changes to FIGO staging. Following the introduction of a new FIGO Staging System, this is usually incorporated into TNM (both UICC and AJCC versions) at a later date. Apart from minor discrepancies in terminology, the UICC and AJCC systems are broadly concurrent.  There are two staging systems for uterine sarcomas. One is to be used specifically for adenosarcomas and the other is for leiomyosarcomas and endometrial stromal sarcomas.1 It is recommended that the latter staging system be used for other malignant uterine mesenchymal neoplasms, such as undifferentiated sarcoma and rhabdomyosarcoma; it is not recommended to provide a pathological stage for STUMP. It is controversial as to whether a pathological stage should be applied to other mesenchymal tumours of uncertain malignant potential which are discussed in this dataset, such as UTROSCT, PEComa and IMT. However, a stage may be applied for those neoplasms which fulfil the criteria for malignancy in the individual tumour types, although this is not mandated.  A tumour should be staged following diagnosis using various appropriate modalities (clinical, radiological, pathological). While the original tumour stage should not be altered following treatment, TNM systems allow staging to be performed on a resection specimen following non-surgical treatment (for example chemotherapy, radiotherapy); in such cases, if a stage is being provided on the pathology report (this is optional), it should be prefixed by ‘y’ to indicate that this is a post-therapy stage.  The reference document TNM Supplement: A commentary on uniform use, 5th Edition (C Wittekind et al. editors) may be of assistance when staging.4  **References**  1 Prat J and Mbatani (2015). Uterine sarcomas. *Int J Gynaecol Obstet* 131 Suppl 2:S105-110.  2 Brierley JD, Gospodarowicz MK and Wittekind C (eds) (2016). *Union for International Cancer Control. TNM Classification of Malignant Tumours, 8th Edition*, Wiley, USA.  3 Amin MB, Edge SB, Greene FL, Byrd DR, Brookland RK, Washington MK, Gershenwald JE, Compton CC, Hess KR, Sullivan DC, Jessup JM, Brierley JD, Gaspar LE, Schilsky RL, Balch CM, Winchester DP, Asare EA, Madera M, Gress DM and Meyer LR (eds) (2017). *AJCC Cancer Staging Manual. 8th Edition*, Springer, New York.  4 Christian Wittekind, James D. Brierley, Anne Lee and Elisabeth van Eycken (eds) (2019). *TNM Supplement: A Commentary on Uniform Use, 5th Edition,* Wiley, USA. | Note that permission to publish the FIGO cancer staging tables may be needed in your implementation. It is advisable to check with FIGO.  Note that permission to publish the TNM cancer staging tables may be needed in your implementation. It is advisable to check.  d Reprinted from Int J Gynaecol Obstet., Volume 131(Suppl 2), Prat J, Mbatani N, Uterine sarcomas, pages S105-10, 2015, with permission from Wiley*.*  e Reproduced with permission. Source: UICC TNM Classification of Malignant Tumours, 8th Edition, eds by James D. Brierley, Mary K. Gospodarowicz, Christian Wittekind. 2016, Publisher Wiley (incorporating any errata published up until 6th October 2020).  f It is recommended that all malignant uterine mesenchymal neoplasms other than adenosarcoma be staged using the staging system for leiomyosarcomas and endometrial stromal sarcomas. |

**Table**

**Table 1: World Health Organization classification of mesenchymal tumours of the uterine corpus.1**

|  |  |
| --- | --- |
| **Descriptor** | **ICD-O codesa** |
| **Mesenchymal tumours specific to the uterus** |  |
| Smooth muscle tumour of uncertain malignant potential (STUMP) | 8897/1 |
| Leiomyosarcoma | 8890/3 |
| Endometrial stromal sarcoma, low grade | 8931/3 |
| Endometrial stromal sarcoma, high grade | 8930/3 |
| Undifferentiated uterine sarcoma | 8805/3 |
| Uterine tumour resembling ovarian sex cord tumour (UTROSCT) | 8590/1 |
| Perivascular epithelioid cell tumour (PEComa) | 8714/3 |
| Inflammatory myofibroblastic tumour | 8825/1 |
| **Mixed epithelial and mesenchymal tumours** |  |
| Adenosarcoma | 8933/3 |
| **Neuroendocrine neoplasia** |  |
| Neuroendocrine tumour NOS | 8240/3 |
| Neuroendocrine tumour, grade 1 | 8240/3 |
| Neuroendocrine tumour, grade 2 | 8249/3 |
| Small cell neuroendocrine carcinoma | 8041/3 |
| Large cell neuroendocrine carcinoma | 8013/3 |
| Carcinoma admixed with small cell neuroendocrine carcinomab | 8045/3 |
| Carcinoma admixed with large cell neuroendocrine carcinomab | 8013/3 |

a These morphology codes are from the International Classification of Diseases for Oncology, third edition, second revision (ICD-O-3.2).3 Behaviour is coded /0 for benign tumours; /1 for unspecified, borderline, or uncertain behaviour; /2 for carcinoma in situ and grade III intraepithelial neoplasia; /3 for malignant tumours, primary site; and /6 for malignant tumours, metastatic site. Behaviour code /6 is not generally used by cancer registries. Incorporates all relevant changes from the 5th Edition Corrigenda June 2021.2

b This terminology is synonymous with the ICD-0 terminology of combined small/large cell neuroendocrine carcinomas.

© World Health Organization/International Agency for Research on Cancer. Reproduced with permission.

**References**

1 WHO Classification of Tumours Editorial Board (2020). *Female Genital Tumours, WHO Classification of Tumours, 5th Edition, Volume 4*. IARC Press, Lyon.

2 WHO Classification of Tumours Editorial Board (2021). *Female Genital Tumours, WHO Classification of Tumours, 5th Edition, Volume 4 - Corrigenda June 2021*. Available from: https://publications.iarc.fr/Book-And-Report-Series/Who-Classification-Of-Tumours/Female-Genital-Tumours-2020 (Accessed 16th June 2021).

3 Fritz A, Percy C, Jack A,  Shanmugaratnam K, Sobin L, Parkin DM  and Whelan S (eds) (2020). *International Classification of Diseases for Oncology, Third edition, Second revision ICD-O-3.2*. Available from: http://www.iacr.com.fr/index.php?option=com\_content&view=category&layout=blog&id=100&Itemid=577 (Accessed 16th June 2021).